

GENOTYPE ENVIRONMENT ANALYSES of COTTON CULTIVAR TRIALS

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Technical Report

to

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PREFACE

This report follows on from that of Williams and van Ewijk.

We have followed the same format to that of Williams and van Ewijk in the naming of years, locations, and cultivars, except where indicated.

Data for 1984/85 and 1985/86 are included in our analysis making a total of 12 years data.

1. DESCRIPTION of PROJECT

1.1 Outline

The purpose of this project is the same as that of Williams and van Ewijk, namely

to conduct a critical examination of the ACCT data from 1974/75 to 1985/86 and to learn more about the relative behaviour of cultivars over the testing sites and the effectiveness of locations to evaluate cultivar differences.

We have concentrated on the numerical classification approach to analysis of the data (Byth, Eisemann and DeLacy, 1976).

1.2 Cultivars

The 108 cultivars tested in the 12 years of trial are listed in Table 1.2.1. We have allocated a "2-symbol" abbreviation to facilitate the labelling of graphs.

Unlike ideal genotype x location x year studies where the genotypes remain the same over the course of the study, the data for this study are taken from a practical breeding program where a large number of the genotypes under test were replaced from year to year. Only 52 cultivars were tested in two years or more (Table 1.2.2) and of those only 36 were tested in three years or more.

The data used for this analysis can be divided into two basic groups to facilitate interpretation. In the six years 74/75 to 79/80 most of the genotypes were of USA origin and not particularly well adapted to the Australian environment. Whereas, in the six years 80/81 to 85/86 most of the genotypes were advanced selections from the CSIRO breeding program and better adapted to the Australian environment.

In the trials conducted during 80/81 to 84/85, many of the genotypes being tested were of Deltapine origin (dpl6, dp41, dp55, dp61, sicot1, sicot1f, sicot2, n40-1h, n40-39h, n40-286f, n40-439g, n40-439h). This is significant because trial data from the USA (reference) shows that Deltapine genotypes are more generally adapted to a wider range of environments than other cotton genotypes.

Table 1.2.1 List of cultivars

abbreviation		name	no
16	dp16	deltapine 16	1
sm	dpsm	deltapine sm	2
13	dp13	deltapine sm seln 13	3
ki	kimdell	kimdell 11	4
ho	hopi	hopicala	5
g6	gl06	gl06	6
ms	m8 sok	m8 superokra	7
dc	delcott	delcott 277	8
ta	st7A	stoneville 7A	9
tn	st7AN	stoneville 7A N	10
by	bayou	bayou sm 1	11
ac	acala70	acala 70/185	12
na	namcala	namcala	13
r9	cr79	cr 79	14
r4	cr74	cr 74	15
rx	rex	rex smoothleaf	16
h6	h62	h62-8	17
ha	hancock	hancock	18
m1	mcn1032	macnair 1032	19
t7	st702	stoneville 702	20
b1	ls/obG	ls/ob G090	21
b2	ls/ob	ss/ob s081	22
b3	ls/obs	ls/ob field bulk	23
rp	rivp	riverina poplar	24
rg	rivg	riverina gold	25
61	dp61	deltapine 61	26
37	tamp37	tamcott sp37	27
c4	c4104	coker 4104	28
m8	m8	m8	29
g2	g245	g245-2-11	30
c0	c310	coker 310	31
mv	mv81	mv081	32
70	70-081	70-081-20-12-19-4	33
r2	crl42	cr 142-45	34
gn	st gN	stoneville gl N	35
gf	dp gf	deltapine gl fr	36
r8	crl28	crl28-5	37
c2	c312	coker 312	38
ds	des024	des 024	39
m2	mcn220	macnair 220	40
s2	sicot2	sicot 2	41
4g	40 286f	n40 286 fr	42
s1	sicot1	sicot 1	43
4f	40 286	n40 286 h	44
4d	40 39h	n40 39 h	45
7x	70 28-1	n70 28/1	46
7y	70 28-3	n70 28/3	47
55	dp55	deltapine 55	48
mo	mo63-277	mo63-277 br	49
c6	c600	coker br-600	50
80	dp80	deltapine 80	51
c5	c315	coker 315	52
c1	c511	coker sm 511	53
4k	41 572N	n41 57-2n	54
6i	dp61ipr	deltapine 61-ipr	55
41	dp41	deltapine 41	56

abbreviation	name	no	
7l	dp7l46N	deltapine 7l4 6N	57
4a	40 lh	n40 lh	58
sf	sicotlf	sicot lf	59
s3	sicot3	sicot 3	60
4j	40 439g	n40 439 g	61
4h	40 439h	n40 439 h	62
mo	mo63-277	mo63 277j	63
3a	39 10-4	n39 10/4	64
3b	39 42-8	n39 42/8	65
7z	70 37-10	n70 37/10	66
8a	85 33-8	n85 33/8	67
07	75007	75007-3	68
23	76023	76023-1	69
7b	74 630	n74 630	70
4c	40 39f	n40 39 hf	71
4i	40 439ds	n40 439ds	72
nb	91 4-109	n91 4/109	73
nc	91 7-111	n91 7/111	74
nd	91 8-30	n91 8/30	75
ne	91 9-111	n91 9/111	76
si	siokra	siokra	77
7c	74 480	n74 480	78
6l	dp6l	deltapine 6l-77	79
90	dp90	deltapine 90	80
7d	74 69	n74 69	81
7e	74 250	n74 250	82
9a	98 283	n98 283	83
9b	98 247	n98 247	84
9c	98 249	n98 249	85
9d	98 251	n98 251	86
9e	98 308	n98 308	87
9f	98 320	n98 320	88
9g	98 329	n98 329	89
9h	98 331	n98 331	90
9i	98 335	n98 335	91
9j	98 336	n98 336	92
50	dp50	deltapine 50	93
7f	74 355	n74 355	94
7g	74 720	n74 720	95
7h	74 666	n74 666	96
7i	74 108	n74 666-108	97
4b	40 lh565	lh 565	98
4c	40 lh570	lh 570	99
2a	sic2374	sicot 2-374	100
2b	sic2411	sicot 2-411	101
2c	sic2418	sicot 2-418	102
2d	sic2436	sicot 2-436	103
sn	siokra N	siokra N	104
sb	siokra s	siokra bulk seln	105
ss	siok Ns	siokra N bulk	106
7j	74 720s	n74 720 seln	107
7k	74 355s	n74 355 seln	108

1.3 Locations

We have used "single-letter" abbreviations for location names (Table 1.3.1) to facilitate the labelling of graphs.

Table 1.2.1 List of Locations

abbreviation	location
e	Emerald
b	Biloela
t	Theodore
d	Darling Downs
s	St George
g	Boggabilla
c	Moomin Creek
m	Moree
v	Myall Vale
n	West Namoi
z	Breeza
w	Warren
k	Bourke

The unsprayed locations and Lockyer (see Table 1.2.4 in Williams and van Ewijk) were excluded from our analyses giving a reduced year x location table of 132 cells of which 100 are filled (Table 1.3.2).

This data can be divided into two basic groups to facilitate its interpretation. In the six years 74/75 to 79/80 only eight locations were used for testing giving a year x location table of 48 cells of which 41 are filled. However, eleven locations were used for testing in the years 80/81 to 85/86 giving a year x location table of 66 cells of which 57 are filled. This grouping of the data corresponds to the same grouping of years as determined on genetic origin of cultivars (section 1.2).

Table 1.3.2 Distribution of locations over years

year	location												
	e	b	t	d	s	g	c	m	v	n	z	w	k
74/75		1	1	1	1				1	1		1	
75/76		1	1		1				1	1		1	
76/77		1	1	1	1				1	1		1	
77/78	1	1	1	1	1				1	1		1	
78/79	1	1	1	1	1					1		1	
79/80	1	1	1	1	1				1	1		1	
80/81	1		1	1	1		1	1	1	1		1	
81/82		1	1	1	1			1	1	1		1	
82/83	1	1	1	1	1	1	1	1	1	1		1	
83/84		1		1		1	1	1	1	1		1	
84/85	1	1		1	1	1	1	1	1	1		1	
85/86	1	1	1	1	1	1	1	1	1	1		1	
86/87	1	1		1	1	1		1	1		1	1	1

1.4 Variates

So far only lint yield has been considered in our analyses because this is the variate of most interest to the cotton breeder when genotype-environment interactions are being considered.

1.5 Data

We used the adjusted genotype x location values calculated by Williams and van Ewijk as the starting point of our analyses.

1.6 Sets of data

We derived a number of cultivar x location x year data sets (Table 1.6.1) by combining data over years where cultivars were common (see Table 1.2.2).

Table 1.6.1 Sets of data used in analysis

year	matrix size g * e	year	matrix size g * e
74	18 x 7	74-77	18 x 28
75	25 x 6	74-80	6 x 80
76	25 x 7	80-84	8 x 46
77	25 x 8	80-85	5 x 57
78	16 x 6	82-85	6 x 40
79	16 x 7	83-85	8 x 29
80	25 x 9	85-86	9 x 21
81	25 x 8		
82	25 x 11		
83	25 x 8		
84	30 x 10		
85	16 x 11		
86	16 x 10		

2. METHODS of ANALYSES

2.1 Linear regression analyses

The proportion of genotype-environment information (sum of squares) accounted for by the linear regression coefficients (Finlay and Wilkinson, 1963) is on average 20% (Table 2.1.1) which was too low to help the cotton breeder gain a better understanding of genotype response over environments.

Table 2.1.1 Proportion of ge accounted for by regression

year	R ²	year	R ²
74	27	74-77	8
75	28	74-80	13
76	15	80-84	6
77	23	80-85	6
78	38	82-85	8
79	13	83-85	10
80	26		
81	9		
82	12		
83	11		
84	21		
85	3		
average	19		8

2.2 Pattern analysis

A numerical classification was employed to derive groupings of genotypes and environments following the approach of Byth, Eisemann and DeLacy (1976). This analysis was refined in two ways. Firstly, environments were classified using ge interaction effects as data (DeLacy, 1981). Since differences in environmental mean yield are often large relative to genotypic differences within environments, this ensured that the grouping of environments was sensitive to differential patterns of genotypic performance within environments by removing any overwhelming influence of environmental mean yields. Secondly, objective determination of the number of genotype and environment groups employed in summarizing the data was made using two criteria. These were the size of the final array of genotype and environment groups to be examined, and the amount of variation, particularly that due to ge interactions, which was retained among groups for this array (DeLacy, 1981).

Classification were obtained using an agglomerative, hierarchical numerical classificatory procedure with incremental sum of squares as the clustering strategy and unstandardized squared Euclidean distance as the measure of dissimilarity (Byth et al, 1976). Several supplementary computer programs (Appendix L) were employed to further summarize the results.

2.3 Single year data sets

The number of environments per year ranged from 6 to 11, which was considered small enough not to require truncation of environments but to retain them all as single member environment groups.

Genotypes were classified for each data set and the number of genotype groups truncated at a level whereby the reduced arrays accounted for 55% (or as close as possible) of the ge interaction sum of squares in the original matrices.

The 16 to 30 genotypes in the original data sets were reduced to 3 to 7 genotypes groups (Table 2.3.1). This was an average reduction of 78% in the size of the original matrix, whilst still retaining on average, 84% of the genotype sum of squares and 55% of the ge interaction sum of squares.

Table 2.3.1 Summary of reduced matrices

years	matrix size gen x env	reduced matrix	% reduction matrix	% G SS retained	% GE SS retained
74	18 x 7	4 x 7	78	74	57
75	25 x 6	5 x 6	80	88	57
76	25 x 7	5 x 7	80	84	56
77	25 x 8	6 x 8	76	90	51
78	16 x 6	4 x 6	75	88	63
79	16 x 7	5 x 7	69	85	59
80	25 x 9	6 x 9	76	92	54
81	25 x 8	5 x 8	80	79	52
82	25 x 11	7 x 11	72	88	54
83	25 x 8	4 x 8	84	85	55
84	30 x 10	5 x 10	87	75	57
85	16 x 11	3 x 11	81	75	55
average			78	84	55
74-77	18 x 28	11 x 8	83	95	55
74-80	6 x 50	6 x 5	90	100	58
80-84	8 x 46	8 x 5	89	100	53
80-85	5 x 57	5 x 4	93	100	55
82-85	6 x 40	6 x 5	88	100	61
83-85	8 x 29	8 x 4	86	100	52
average			88		55

2.4 Combined year data sets

Except for the 74-77 data set, the other arrays contained 5 to 8 genotypes which was considered small enough not to require truncation of genotypes but to retain them all as single member genotype groups.

Environments were classified for each data set and the number of environmental groups truncated at a level whereby the reduced arrays accounted for 55% (or as close as possible) of the ge interaction sum of squares in the original matrices.

The 29 to 57 environments in the original data sets were reduced to 4 to 5 environment groups in the reduced arrays (Table 2.3.1). This was an average reduction of 88% in the size of the original array whilst still retaining an average of 55% of the ge interaction sum of squares.

For the 74-77 data set, separate classifications of genotypes and environments were super-imposed to give a two-way reduction of the original 18 x 28 matrix. Two criteria were used to decide the size of the reduced matrix, firstly retaining 55% of the ge interaction sum of squares, and secondly, obtaining the maximum reduction in the size of the original array. The 11 x 8 reduced array was a reduction of 83% in array size, whilst still retaining 95% of the genotype sum of squares and 55% of the ge interaction sum of squares.

3. GENOTYPE RESPONSE to ENVIRONMENTS - SINGLE YEARS

For each single year data set the genotype response to environments was considered by examining:

- (i) the dendogram illustrating the classification of genotype groups (all dendogram figures in Section 3) and the genotype composition of each of those groups (Appendix C).
- (ii) the graph illustrating all the genotype group responses over environments. The overall contribution of genotype main effects and ge interaction effects (Shorter and DeLacy, 1981) is given for the separation of these genotype groups.
- (iii) the graphs illustrating the contrasting responses to environments of the two genotype groups at each fusion level (Appendix J). The contribution of genotype main effects and ge interaction effects is given for each fusion level.
- (iv) the contributions of ge effects to the classification of genotypes at each fusion level (Appendix I).
- (v) the two-way analysis of variance (Appendix A).

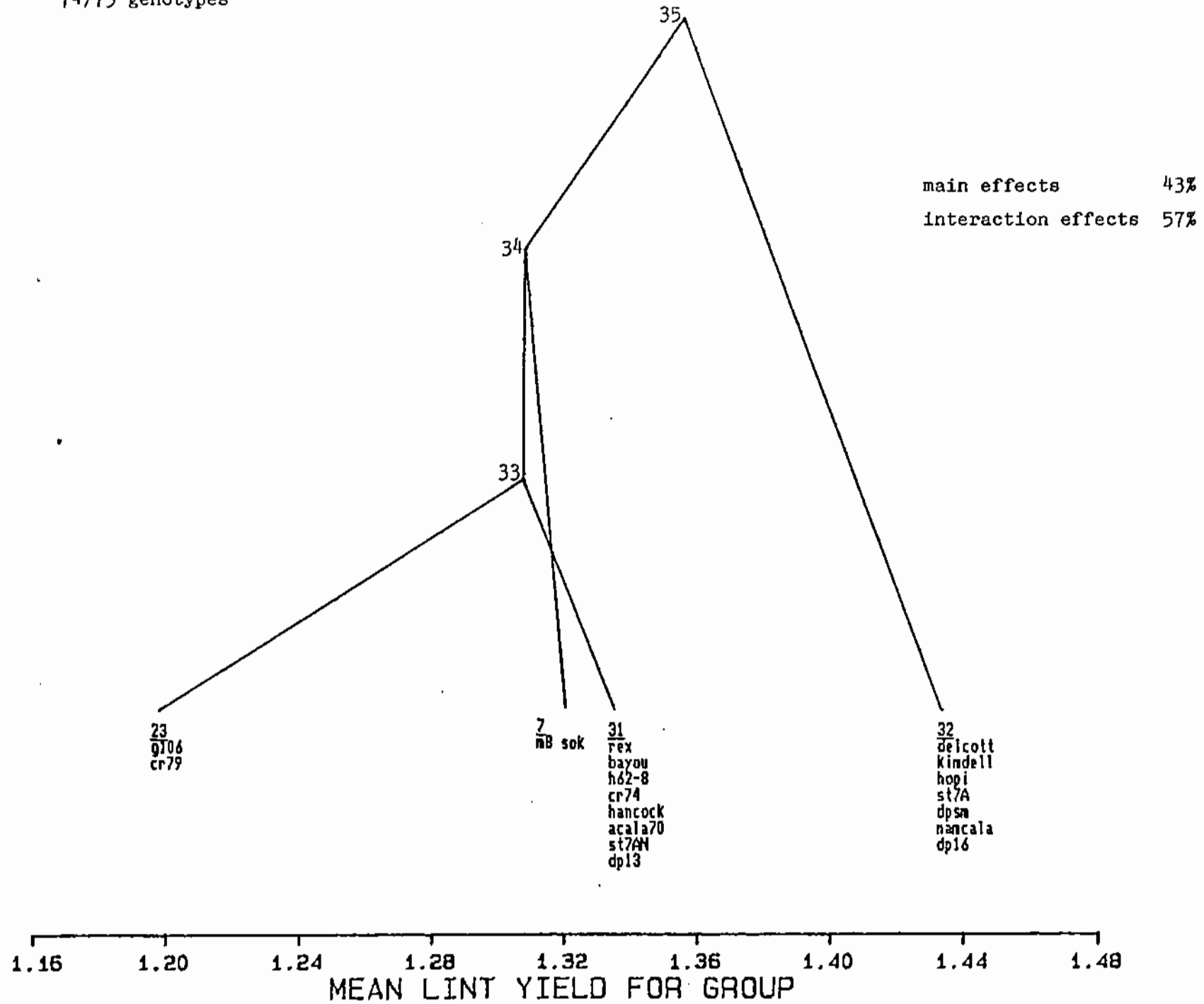
3.1 74/75 genotypes

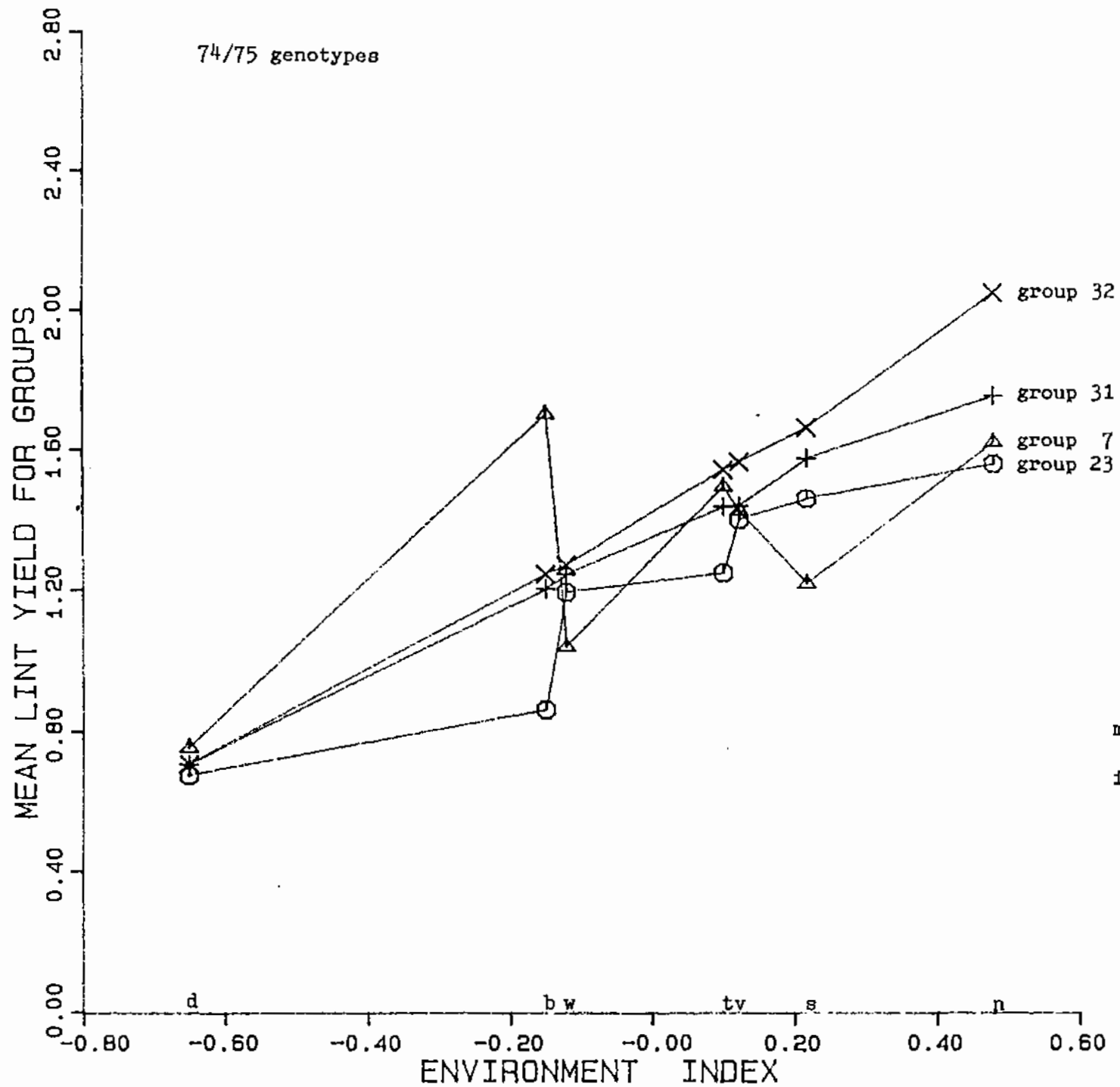
- group 23 - CSIRO bred selections from the Riverina
 - two member group which is low yielding at all environments.
- group 7 - "m8 superokra" has variable response across environments, highest yielding genotype at Biloela and lowest yielding genotype at Warren and St George.
- group 31 - miscellaneous group of genotypes
 - average yield at all environments.
- group 32 - mostly Deltapine related genotypes
 - high yielding genotypes at all locations, except Biloela.

Genotype-environment effects (57%) are a major component in separating genotype groups. The main environments contributing to the separation of genotype groups are

- n separates higher yielding adapted genotypes (group 32) from other genotypes.
- b separates other genotypes into three groups, unadapted (group 23), specifically adapted to biloela (m8sok) and generally adapted (group 31).

74/75 genotypes





main effects 43%

interaction effects 57%

3.2 75/76 genotypes

- group 45 - CSIRO bred selections from the riverina
 - low yielding at all sites.

- group 43 - USA developed selections
 - low to average yields at all locations except Warren where it is low yielding.

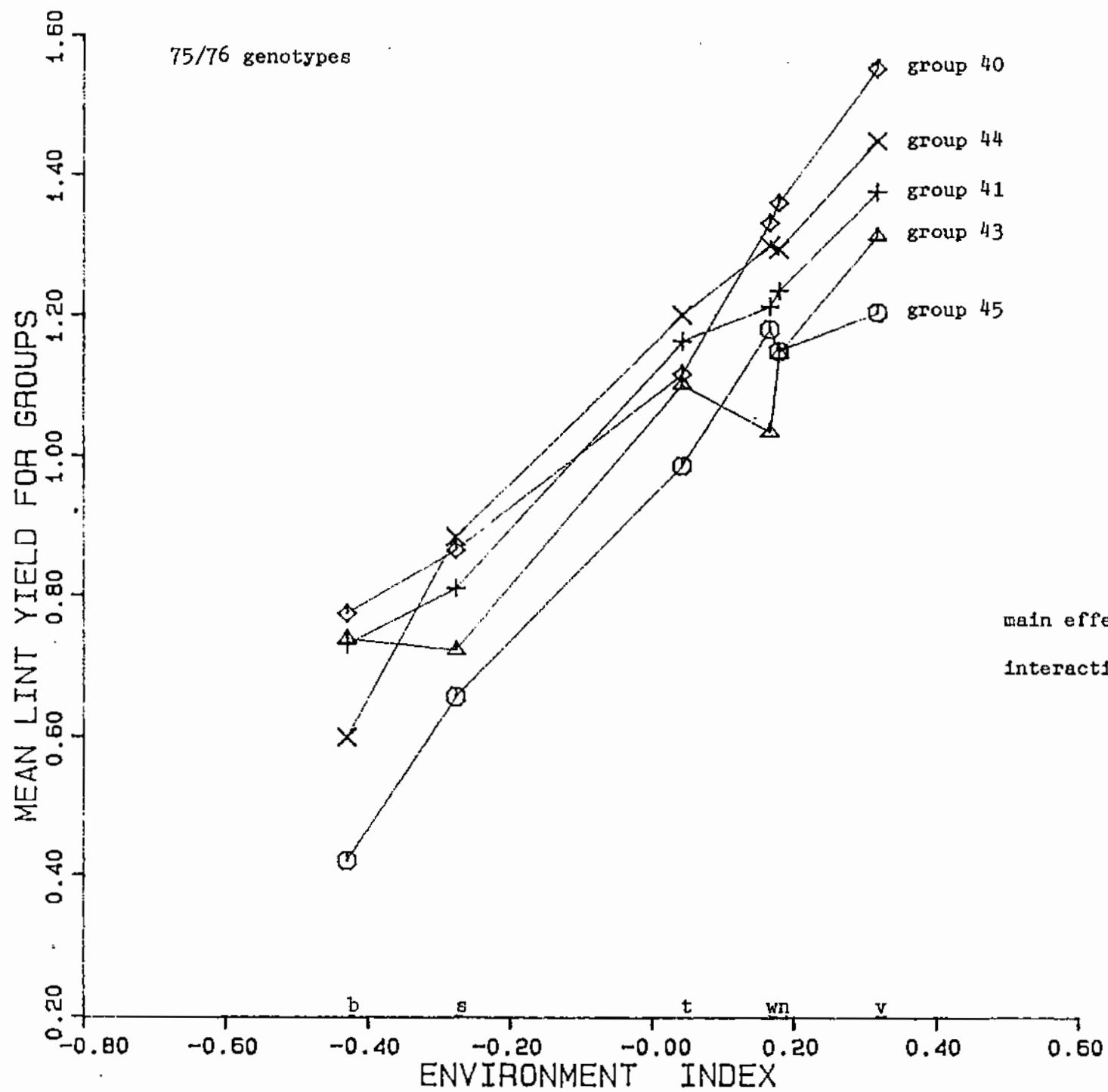
- group 41 - stoneville related genotypes
 - average yields at all locations.

- group 44 - mostly USA developed genotypes
 - high yielding group at most sites.

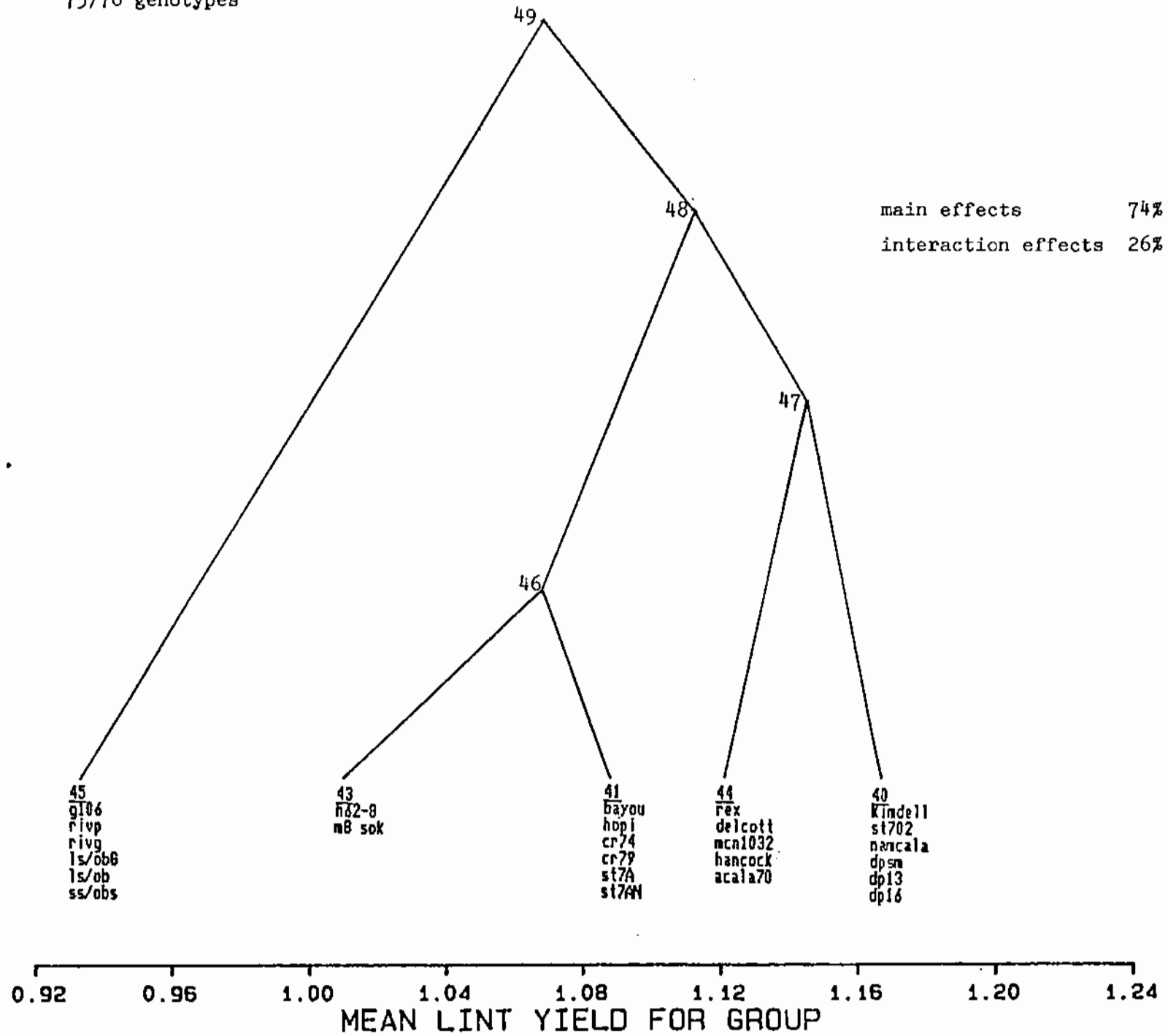
- group 40 - recently developed varieties of USA origin
 - highest yielding group at all sites except Theodore.

Genotype groups are mostly separated by genotype main effects (74%) and not ge interaction effects. The main environments contributing to the separation of genotype groups are

b,v main locations for separating genotype groups



75/76 genotypes



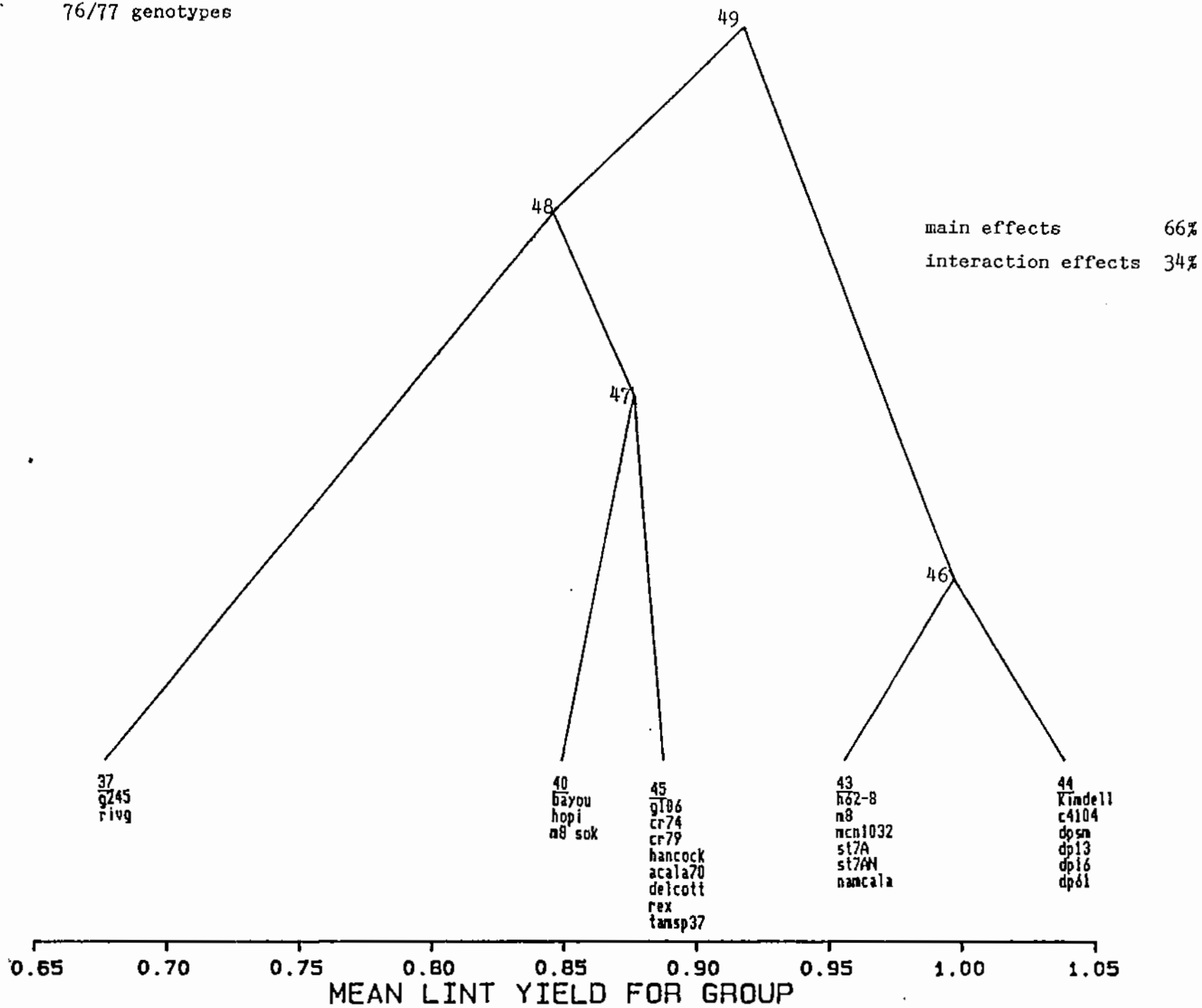
3.3 76/77 genotypes

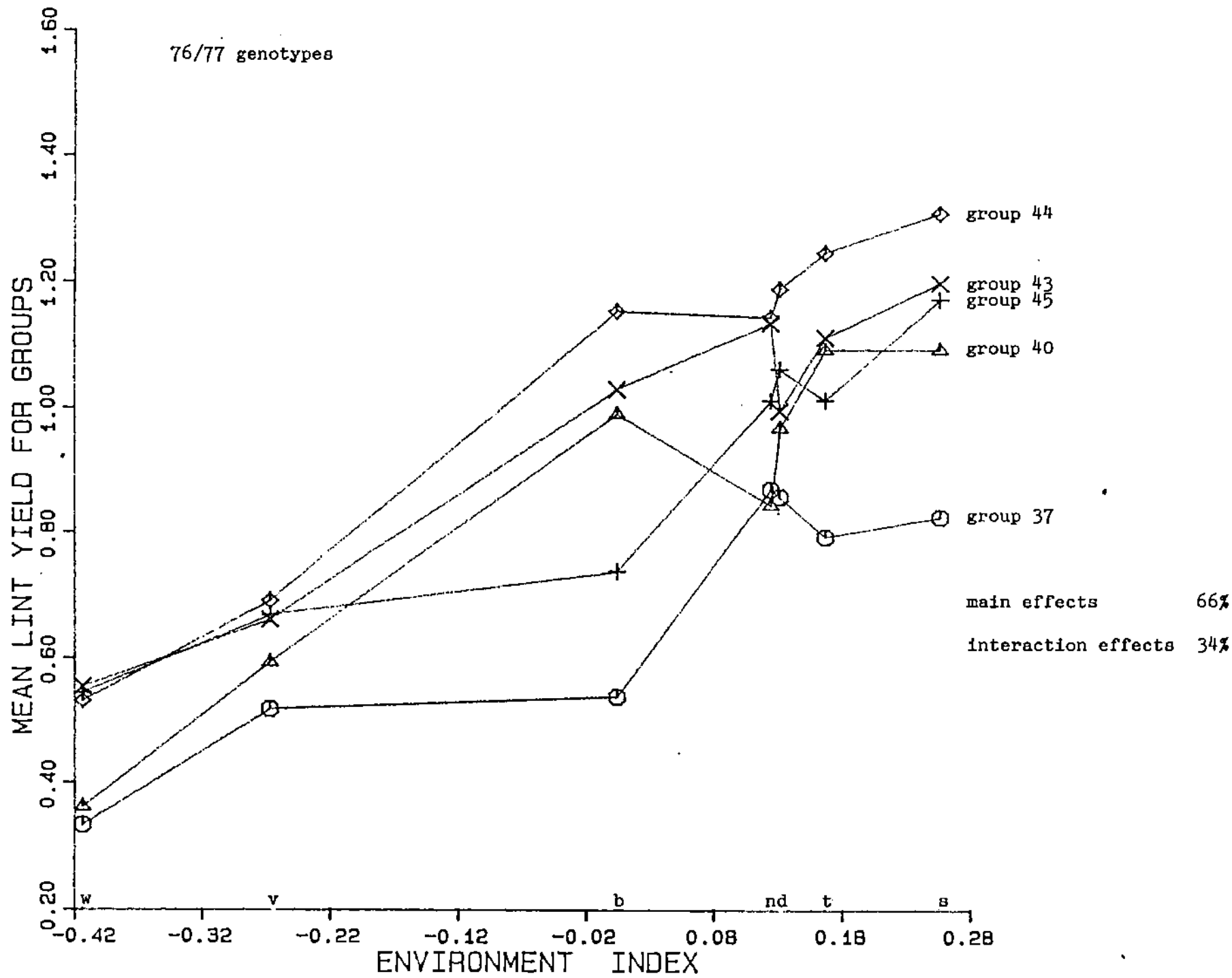
- group 37 - CSIRO bred selections from the Riverina
 - group of two genotypes with low yields at all sites.
- group 40 - USA developed genotypes
 - low to average yielding group.
- group 45 - miscellaneous group of genotypes
 - average yielding group with low yields at high yielding environments.
- group 43 - mostly Stoneville related genotypes
 - high to average yields at all locations except Darling Downs which is low yielding.
- group 44 - deltapine related genotypes
 - high yielding at all environments.

Genotype groups are separated by both genotype main effects (64%) and genotype x environment interaction effects (34%). The main environments contributing to the separation of genotype groups are

- b main environment for separating genotypes into five groups
- w contrasting environment to biloela for separating groups 40 and 45.

76/77 genotypes





3.4 77/78 genotypes

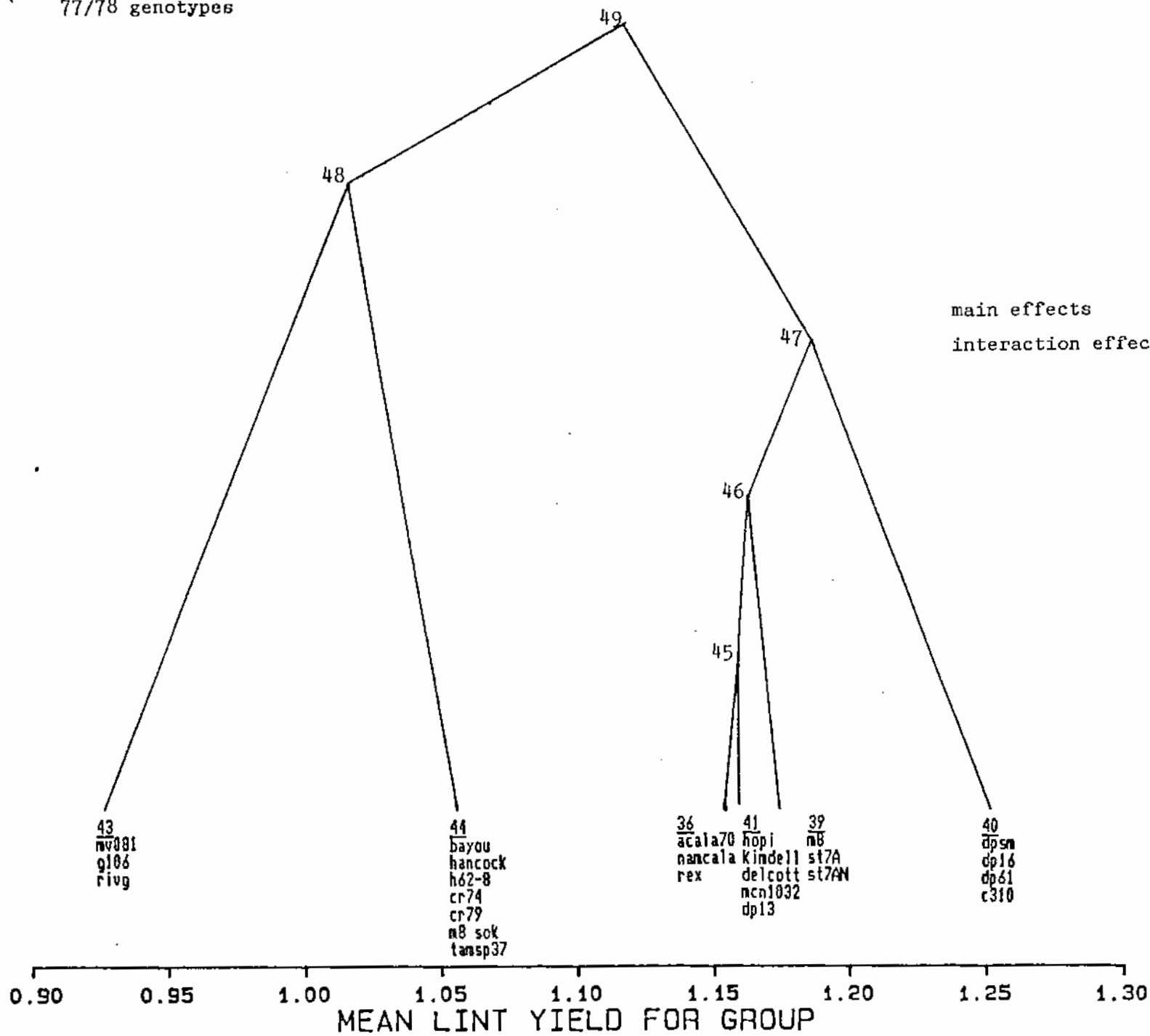
- group 43 - CSIRO developed selections from Riverina
 - low yielding at all sites.
- group 44 - USA developed genotypes which are low to average yielding at all locations.
- group 36 - acala related genotypes.
 - average yield at most locations but relative high yielding at Darling Downs and low yielding at Biloela.
- group 41 - mixed group of genotypes.
 - average yield at all locations.
- group 39 - stoneville relative genotypes.
 - average yield at most locations, but high yielding at Biloela and West Namoi and relatively low yielding at Warren and Theodore.
- group 40 - recently developed varieties of USA origin.
 - high yielding at all locations except st george.

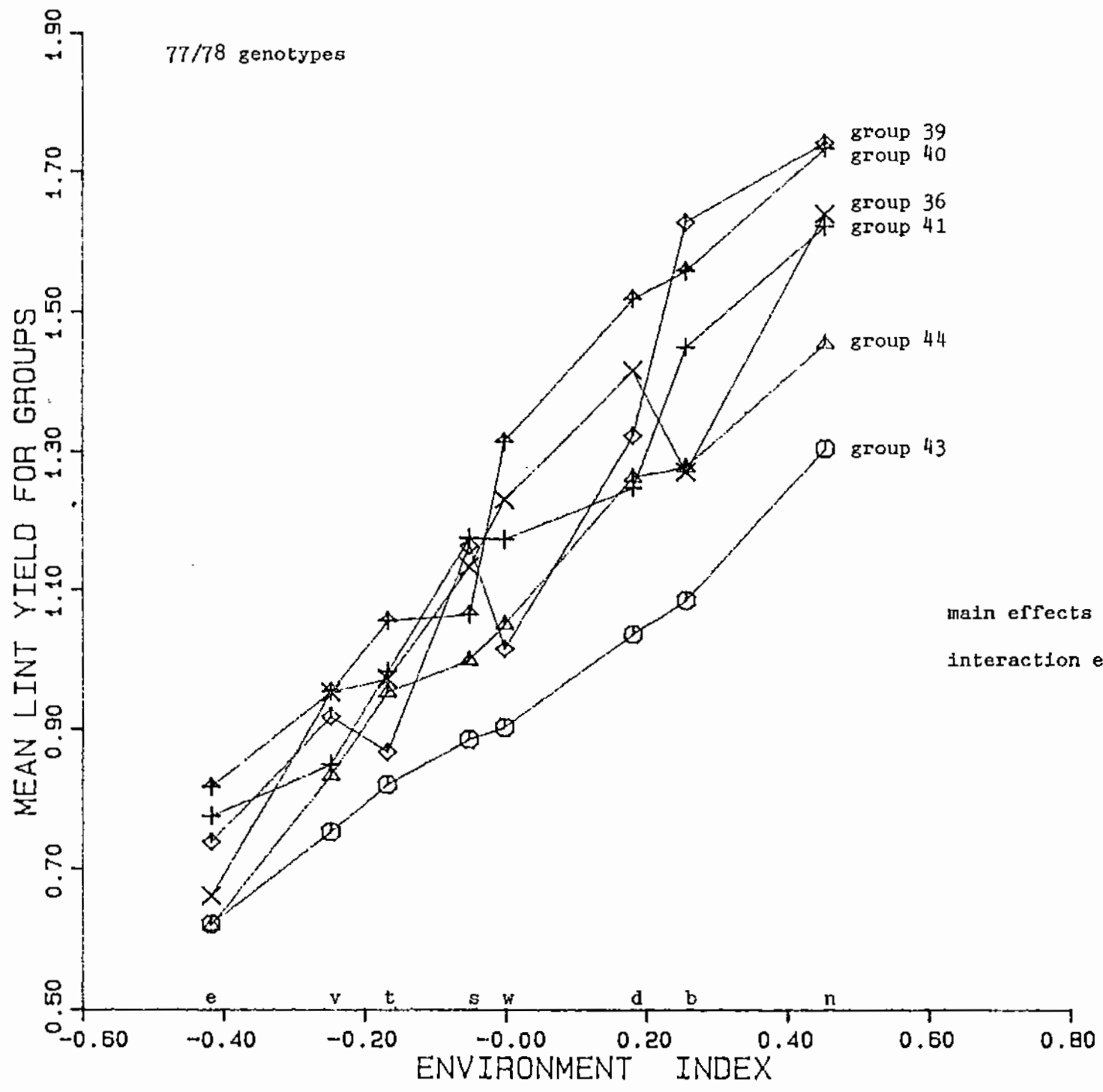
Groups 36, 39 and 41 have the same average yields but they differ in their response over environments.

Both genotype main effects (68%) and ge interaction effects (32%) contribute to the separation of these genotype groups. The main environments contributing to the separation of genotype groups are

- b,n,d,w four main locations for separating genotypes into four groups, unadapted (group 43), low to medium yield (group 44), medium to high yield (group 46), and high yield (group 40).
- b contrasting environment to either warren or darling downs for separating three groups, acala related genotypes (group 36), stoneville related genotypes (group 39), and group 41 (mixed group of genotypes).

77/78 genotypes





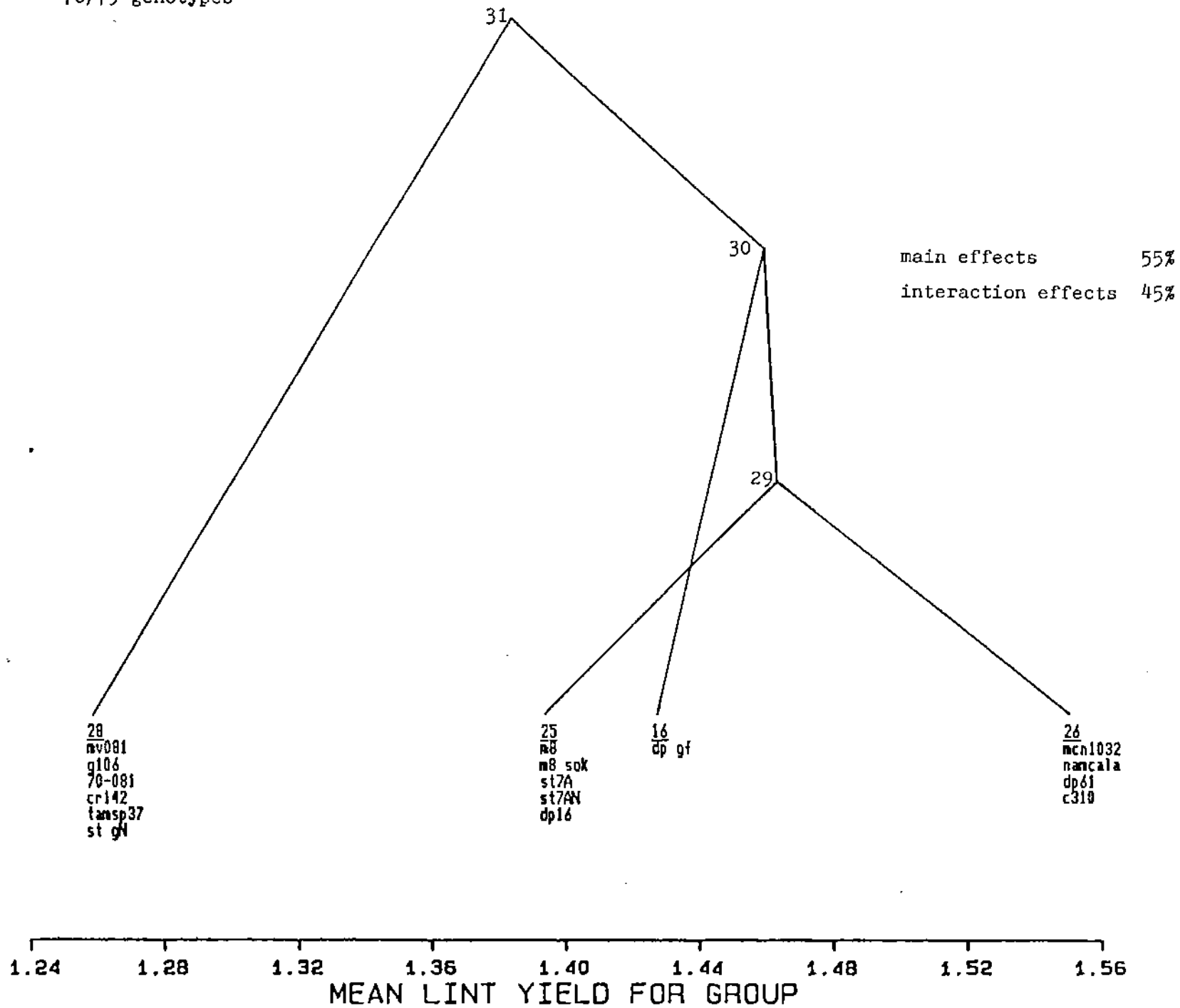
3.5 78/79 genotypes

- group 28 - mostly CSIRO selections developed for the Riverina
 - low yielding at all locations.
- group 25 - stoneville related genotypes
 - average yield at all locations.
- group 16 - "deltapine glabrous frego" has variable yield response over environments, highest yielding genotype at some sites and lowest yielding genotype at other sites.
- group 26 - recently developed varieties of USA origin.
 - high yielding at all locations, except emerald.

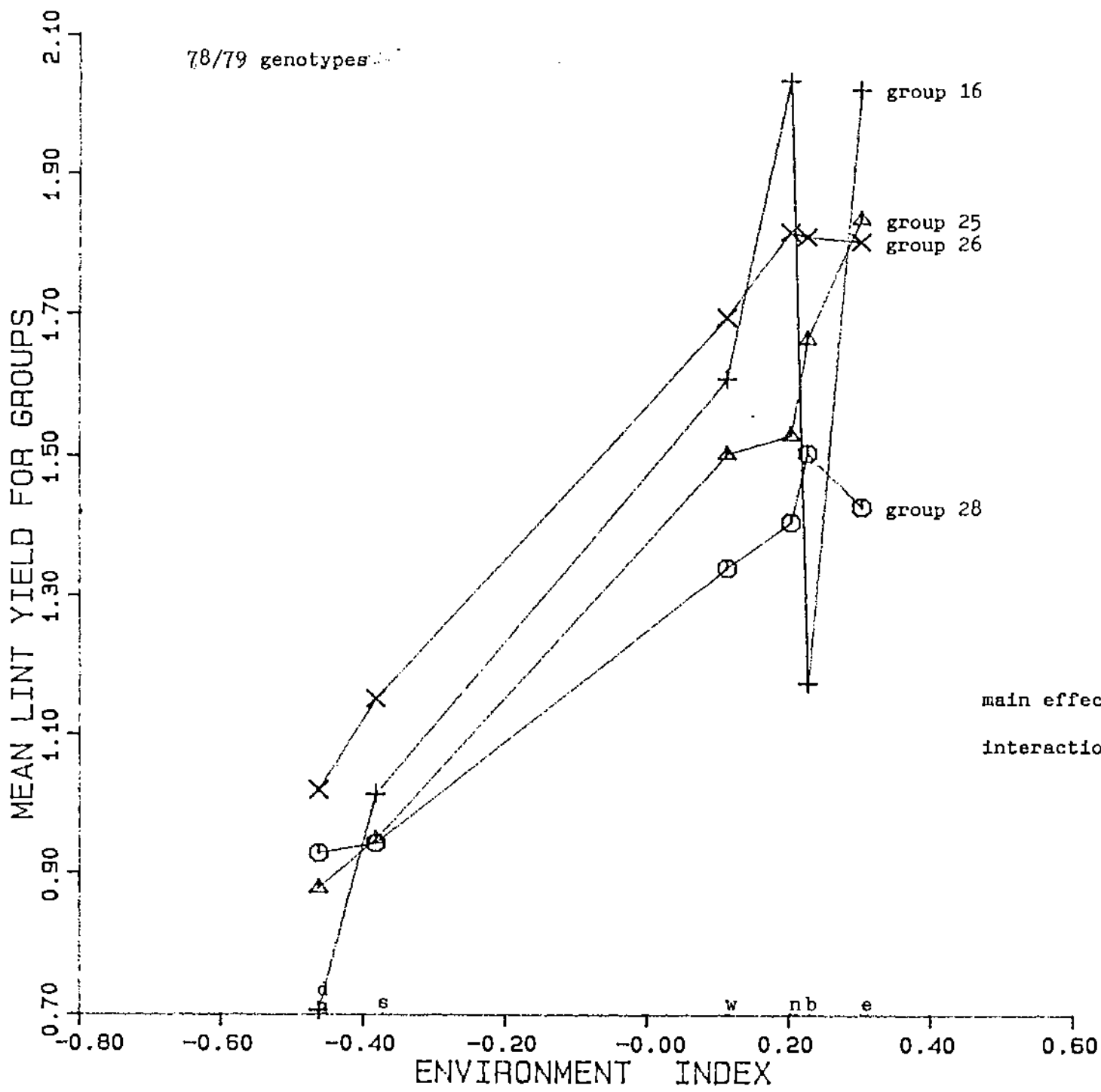
Both genotype main effects (55%) and ge interaction effects (45%) contribute to the separation of genotype groups. The main environments contributing to the separation of genotype groups are

- e separates unadapted genotypes (group 28) from the rest
- n separates Stoneville related genotypes (group 25) from recently developed USA cultivars (group 26)
- b separates "deltapine gl fr" from other genotypes

78/79 genotypes



78/79 genotypes



main effects 55%
interaction effects 45%

3.6 79/80 genotypes

- group 2 - "gl06" is CSIRO selection from Riverina
 - low yielding at all sites.

- group 14 - "crl26" is CSIRO selection from Riverina
 - low to average yield
 - variable response over environments, similar "dp gf".

- group 27 - stoneville related genotypes.
 - average yield at all sites.

- group 13 - "dp gf" has variable yield response over environments, highest yielding genotype at some sites and lowest yielding genotype at other sites.

- group 26 - recently developed varieties of deltapine and coker origin.
 - high yields at all environments.

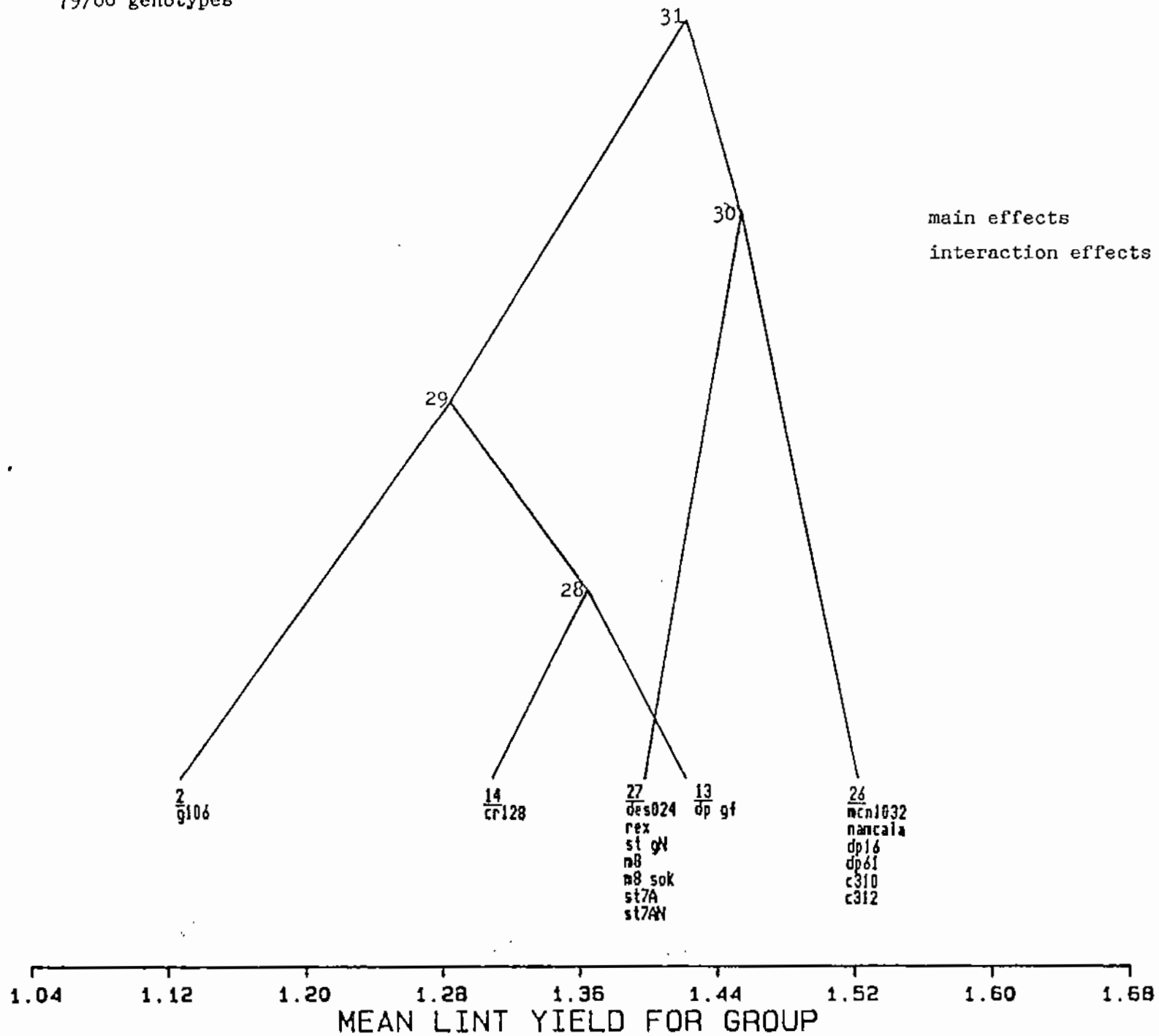
Three genotypes, the single member groups 2, 13, 14, make the largest contribution to ge interaction.

Both genotype main effects (57%) and ge interaction effects (43%) contribute to the separation of genotype groups. The main environments contributing to the separation of genotype groups are

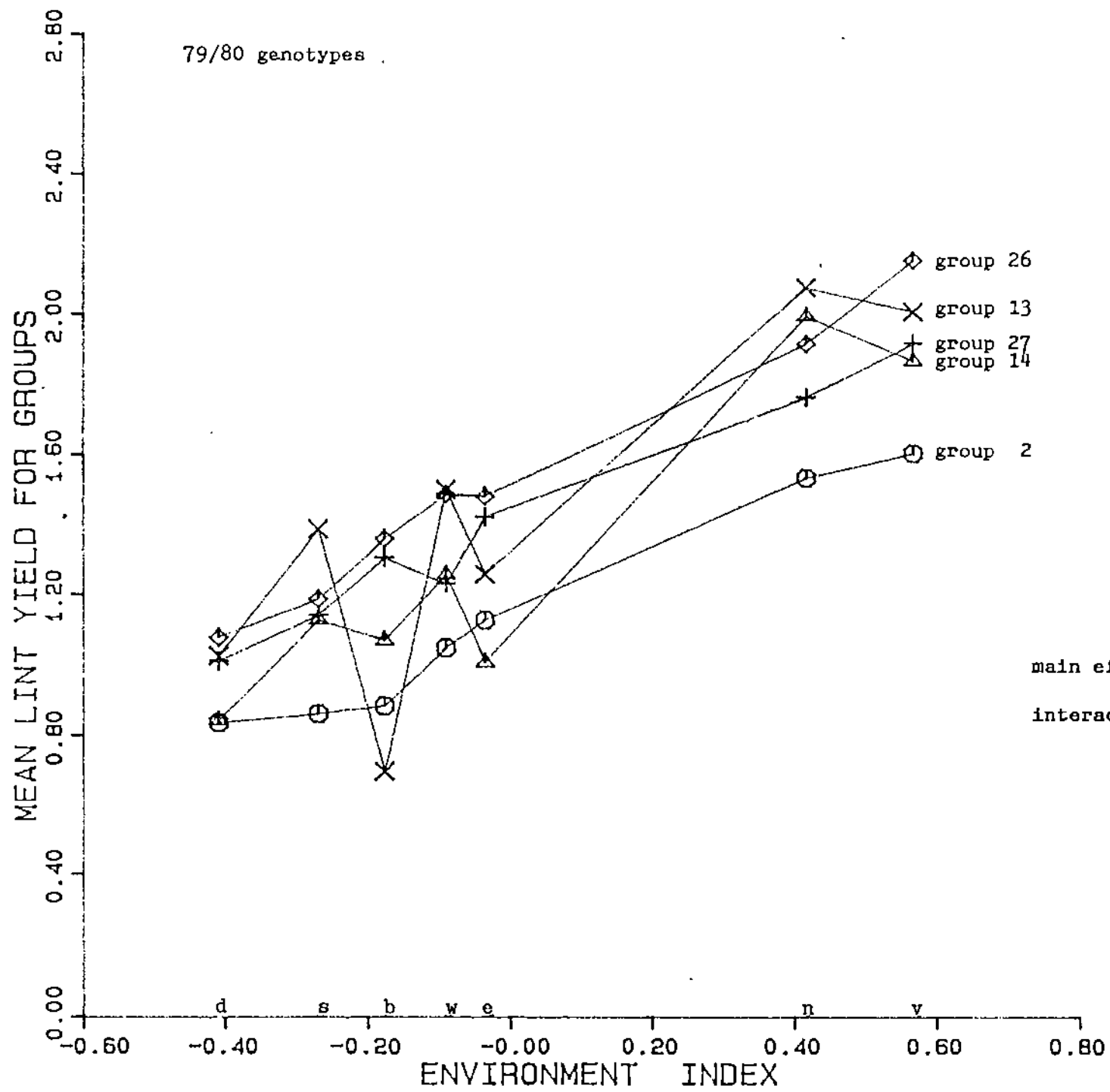
w,v separates stoneville related genotypes (group 27) from deltapines and cokers (group 26)

b separates "deltapine gl fr" from other genotypes

79/80 genotypes



79/80 genotypes



3.7 80/81 genotypes

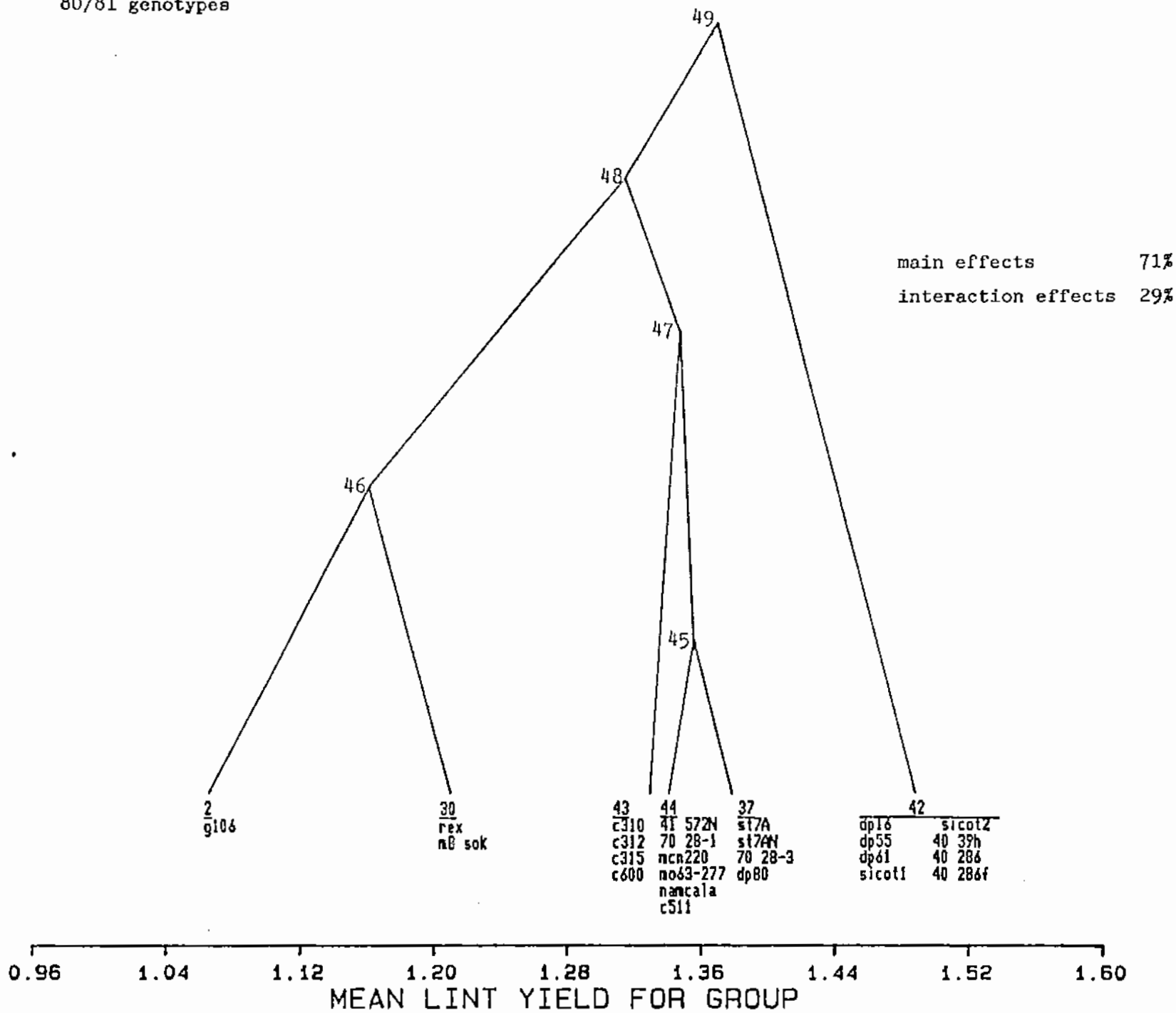
- group 2 - "gl06" is CSIRO selection from Riverina
 - lowest yielding genotype at all sites.
- group 30 - two member group of USA origin.
 - second lowest yielding group at all environments.
- group 43 - coker varieties.
 - average yielding at all sites except high yielding at theodore and low yielding at warren.
- group 44 - mixed group of genotypes of earlier maturity.
 - average yield at all environments.
- group 37 - stoneville related genotypes.
 - average yield at all locations except high yield at Moree.
- group 42 - deltapine related genotypes.
 - highest yielding at all locations, except theodore.

Group 43 (coker varieties), group 44 (early maturity), and group 37 (stoneville related genotypes) have similar mean yields but differ in their response over environments.

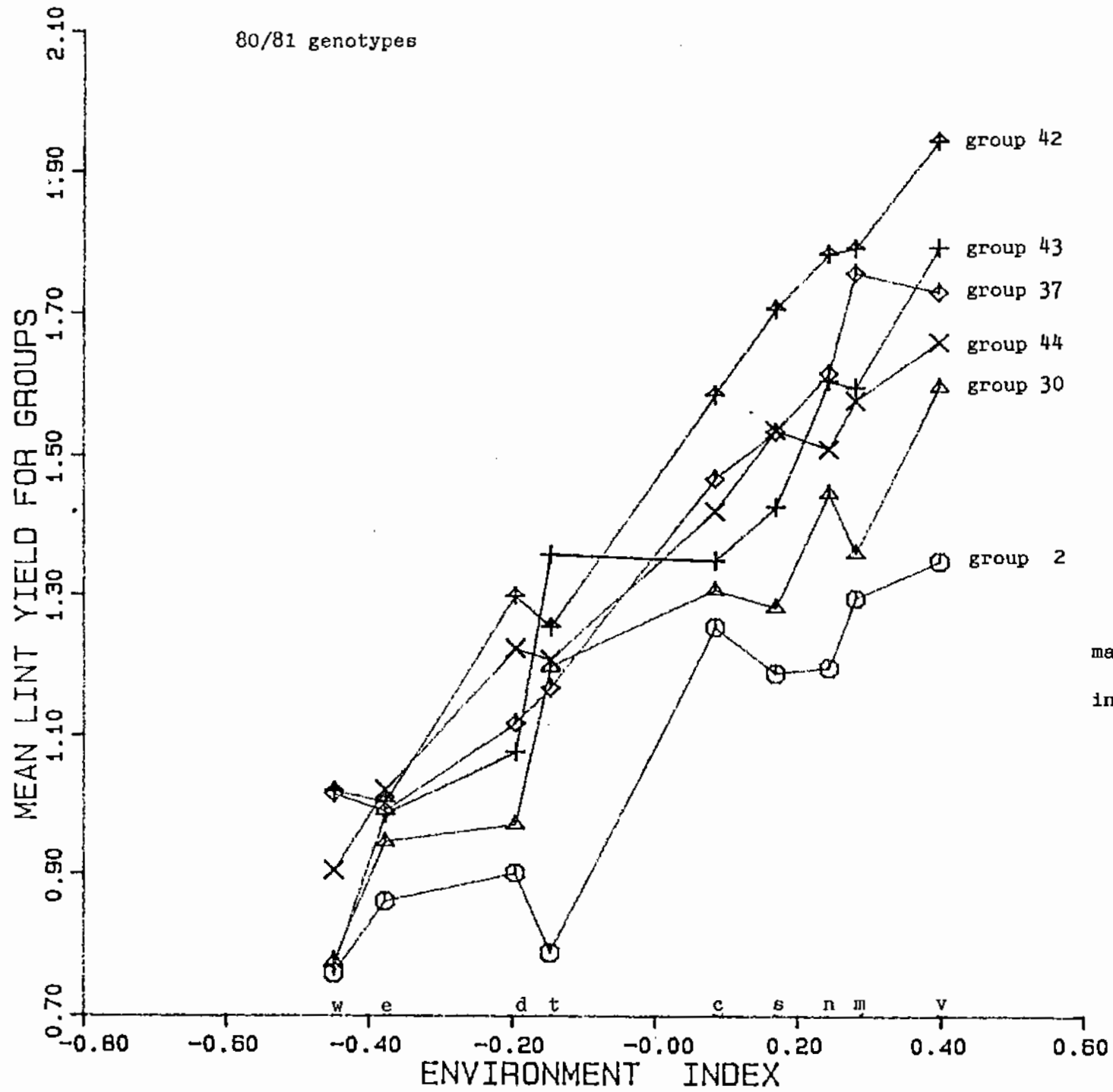
Genotype groups are mostly separated by genotype main effects (71%) and not ge effects. The main environments contributing to the separation of genotype groups are

- v,n,s separates Deltapines (group 42) from other genotypes
- m separates low yielders (gl06, rex, m8) from Coker and Stoneville related genotypes (group 47)
- t separates Cokers (group 43) from other genotypes
- m/d two contrasting locations for separating stoneville genotypes (group 37) from McNair, Namcala, and Coker genotypes (group 44)

80/81 genotypes



80/81 genotypes



3.8 81/82 genotypes

- group 42 - namcala related genotypes.
 - low yielding at all sites except Darling Downs.
- group 43 - early maturity genotypes.
 - low to average yield at all sites except darling downs.
- group 45 - deltapine related genotypes plus other CSIRO selections.
 - average yield at all locations except low at darling downs.
- group 40 - deltapine related genotypes plus coker 315.
 - average yield at all locations except high at warren.
- group 44 - deltapine related genotypes.
 - high yielding at all locations except darling downs.

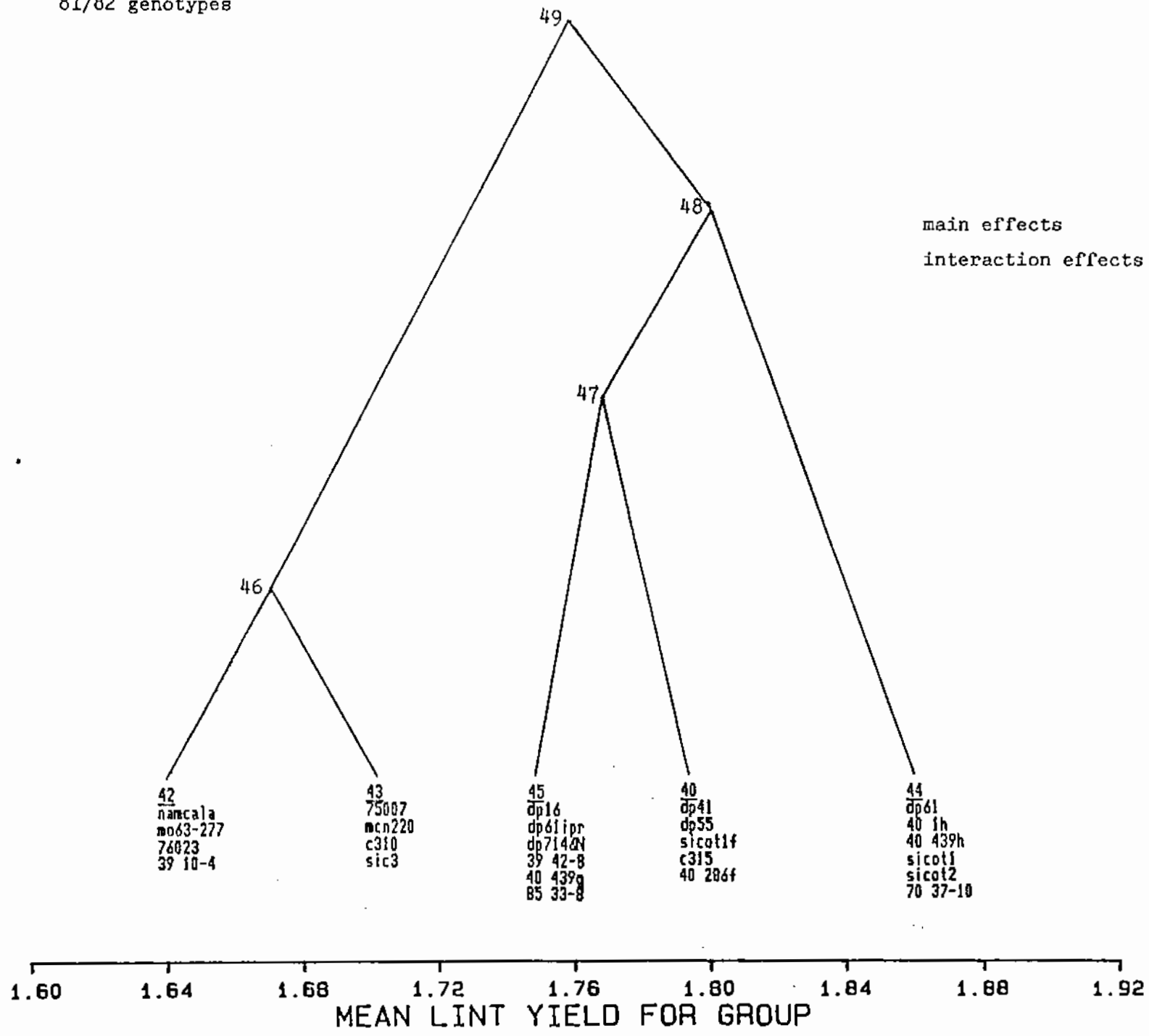
The Deltapine related genotypes (groups 40, 44, 45) fuse to form group 48 which is different from the non-Deltapine related genotypes (groups 42, 43) which fuse to form group 46.

Genotype-environment interaction effects (51%) are a major component in the separation of genotype groups. The main environments contributing to the separation of genotype groups are

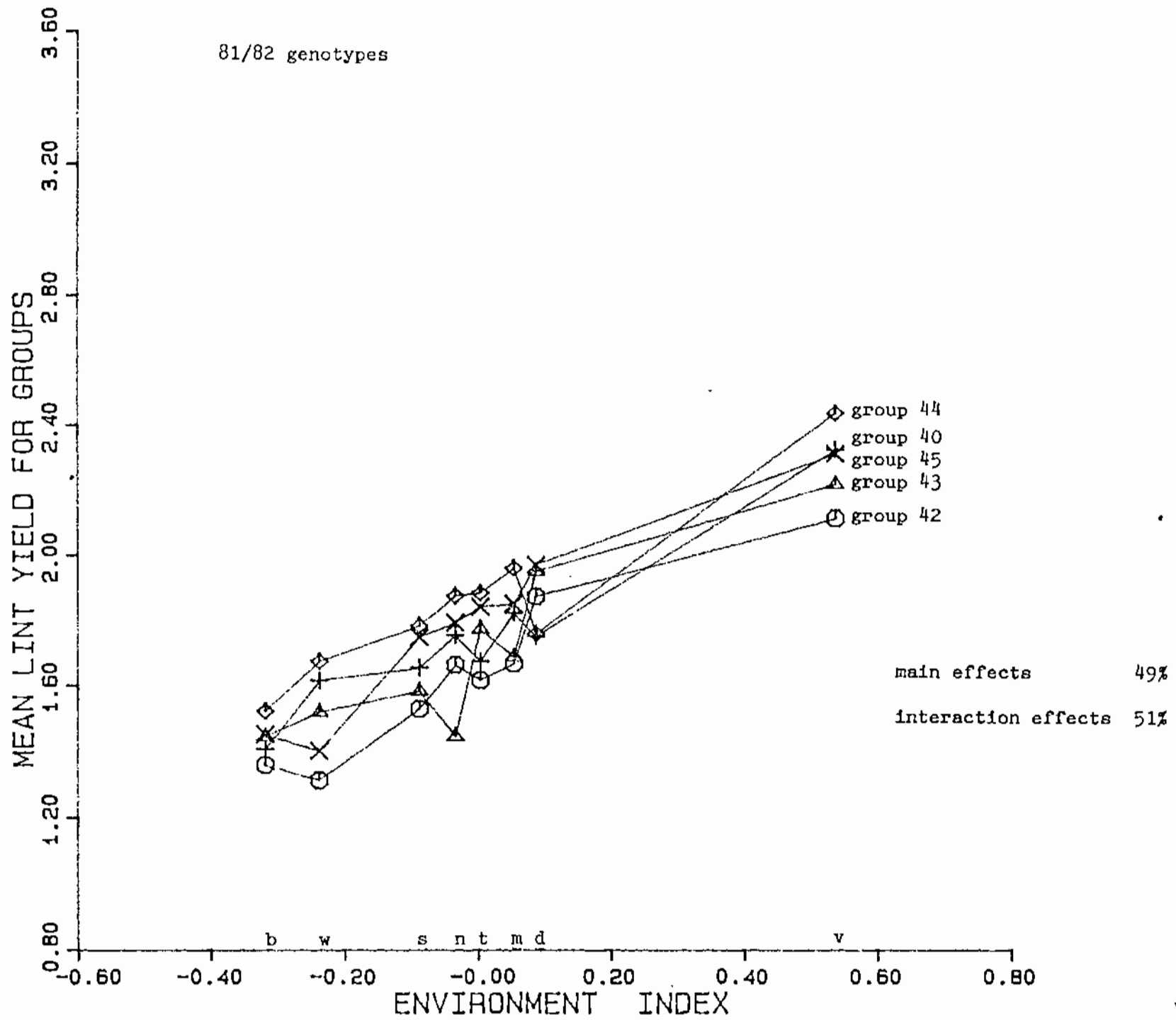
- n,m,v/d separates deltapine related genotypes (group 48) from short season genotypes (group 46)
- w/d two contrasting locations for separating deltapine genotypes into three response groups (group 40,44,45)

Myall vale has a higher yield than all other locations.

81/82 genotypes



81/82 genotypes



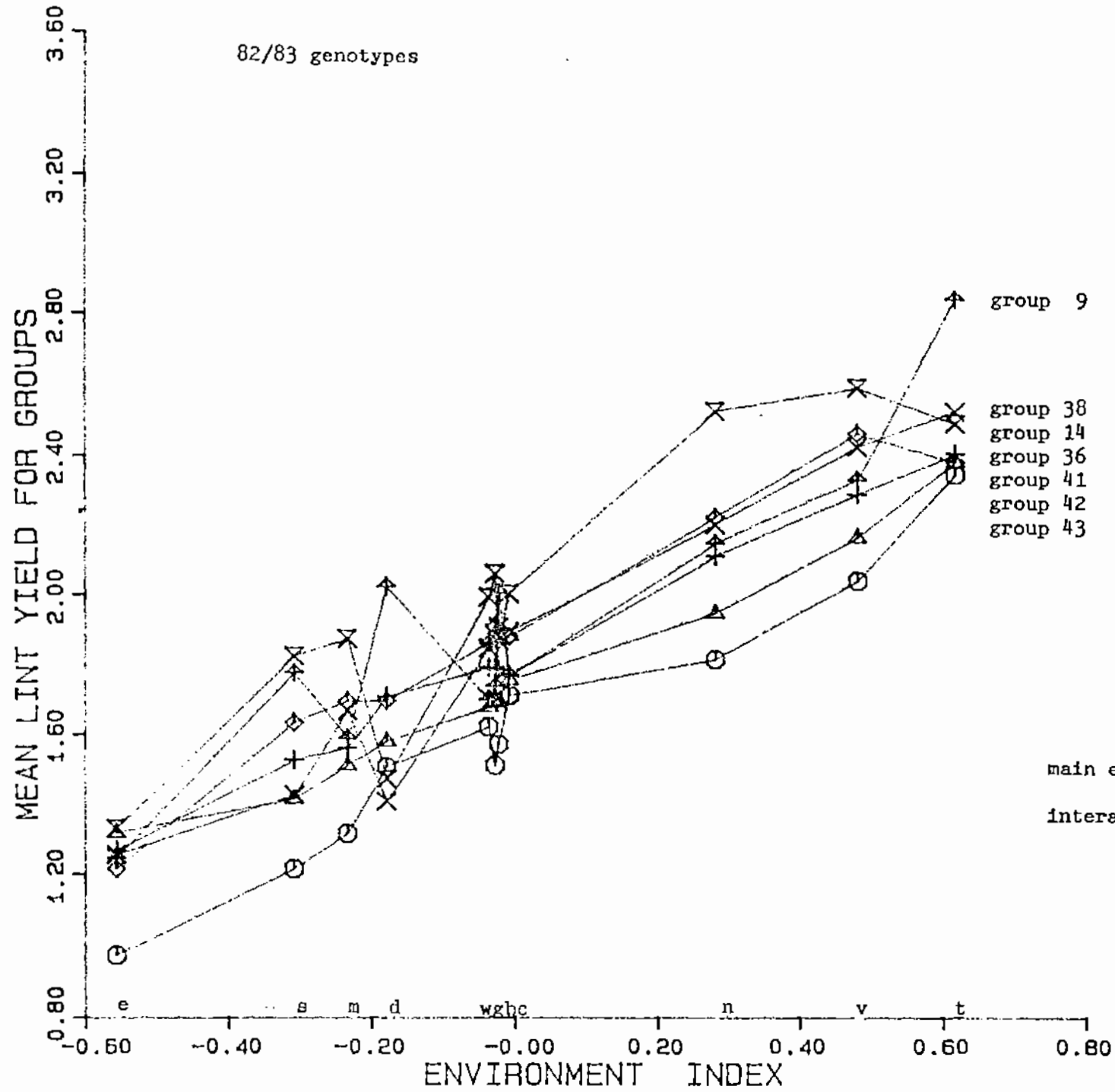
3.9 82/83 genotypes

- group 43 - early maturity genotypes.
 - lowest yielding group at all locations except darling downs.
- group 42 - mixed group of genotypes which are low yielding at most locations.
- group 41 - mostly Deltapine related genotypes of average yield.
- group 38 - genotypes which are low yielding in southern Queensland.
- group 36 - deltapine related genotypes of above average yield.
- group 9 - "coker 315" is high yielding at Queensland locations and average at NSW locations.
- group 14 - "n40 439h" is highest yielding at most sites except darling downs, biloela and theodore.

Both genotype main effects (58%) and ge interaction effects (41%) contribute to the separation of genotype groups. The main environments contributing to the separation of genotype groups are

- n,v separates deltapine related genotypes (group 48) from other genotypes (group 47)
- d separates deltapine genotypes into groups which have general adaptation or specific adaptation
- t separates Coker (group 9) from other genotypes
- e separates short season cultivars (group 43) from other genotypes

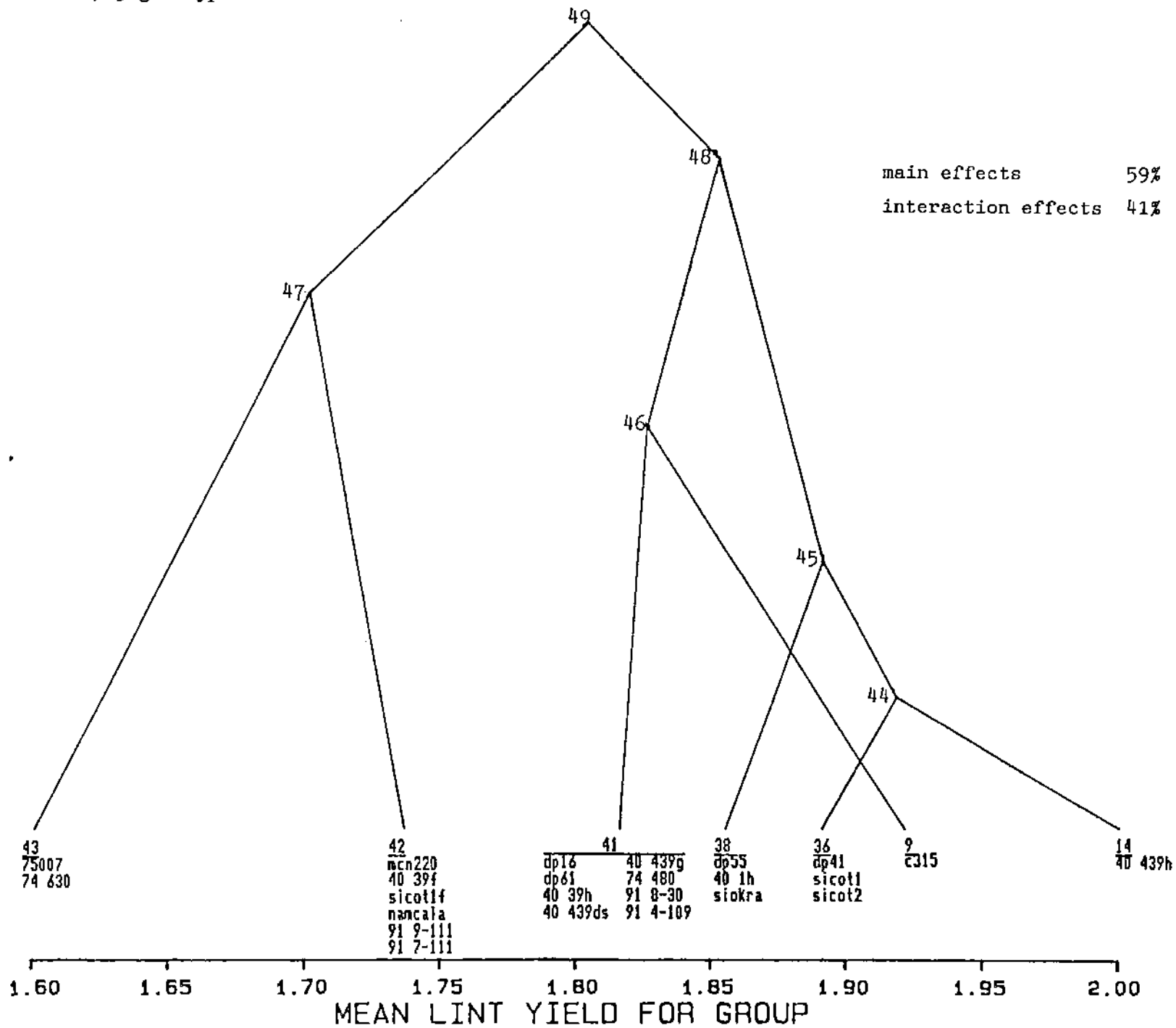
82/83 genotypes



main effects 59%

interaction effects 41%

82/83 genotypes



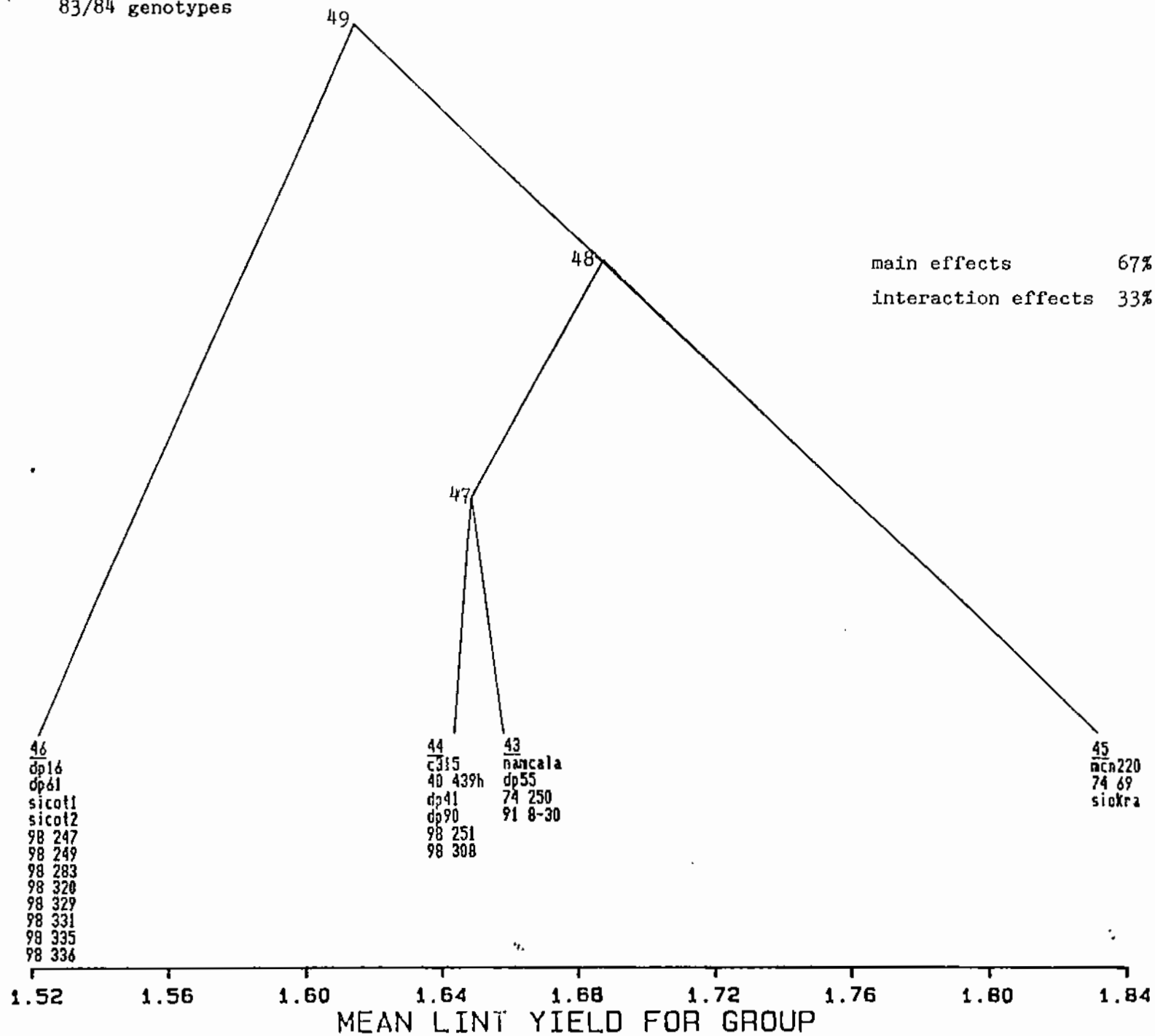
3.10 83/84 genotypes

- group 46 - deltapine related genotypes.
 - low yielding at all sites except central Queensland.
- group 44 - mixed group of genotypes of average yield.
- group 43 - namcala related genotypes.
 - average yield at all sites except Biloela where it is low.
- group 45 - early maturity genotypes.
 - high yielding at all sites except central Queensland.

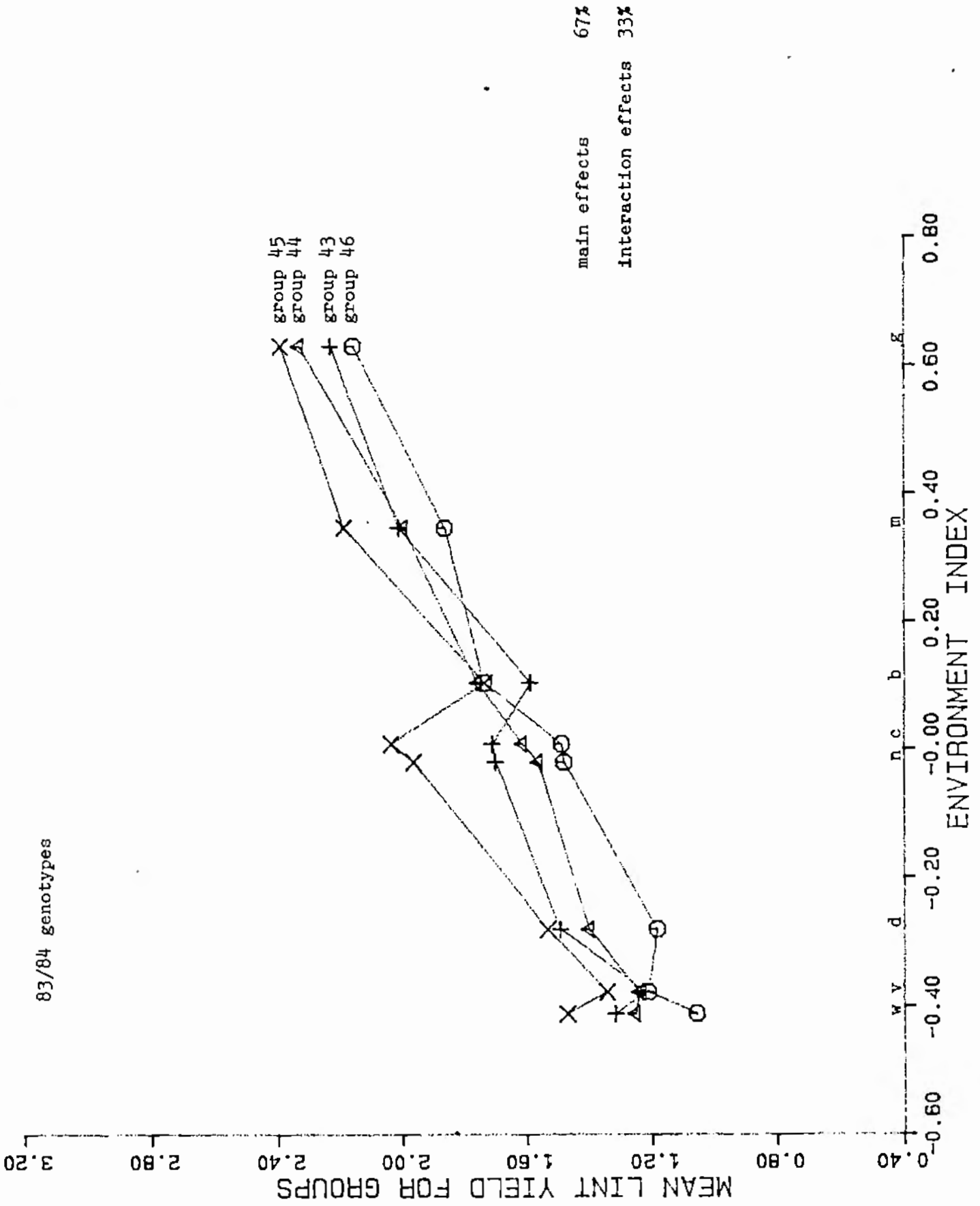
Groups 45, 46 and 47 are mostly separated by genotype main effects (67%). Groups 43 and 44 are separated by ge interaction effects, with biloela and west namoi being the two contrasting environments. The main environments contributing to the separation of genotype groups are

- d,w,c,n separates genotypes into three groups, short season genotypes (group 45), n98 and deltapine genotypes (group 46), and Namcala, Coker genotypes (group 47)
- b separates namcala related genotypes (group 43) from other genotypes

83/84 genotypes



83/84 genotypes



3.11 84/85 genotypes

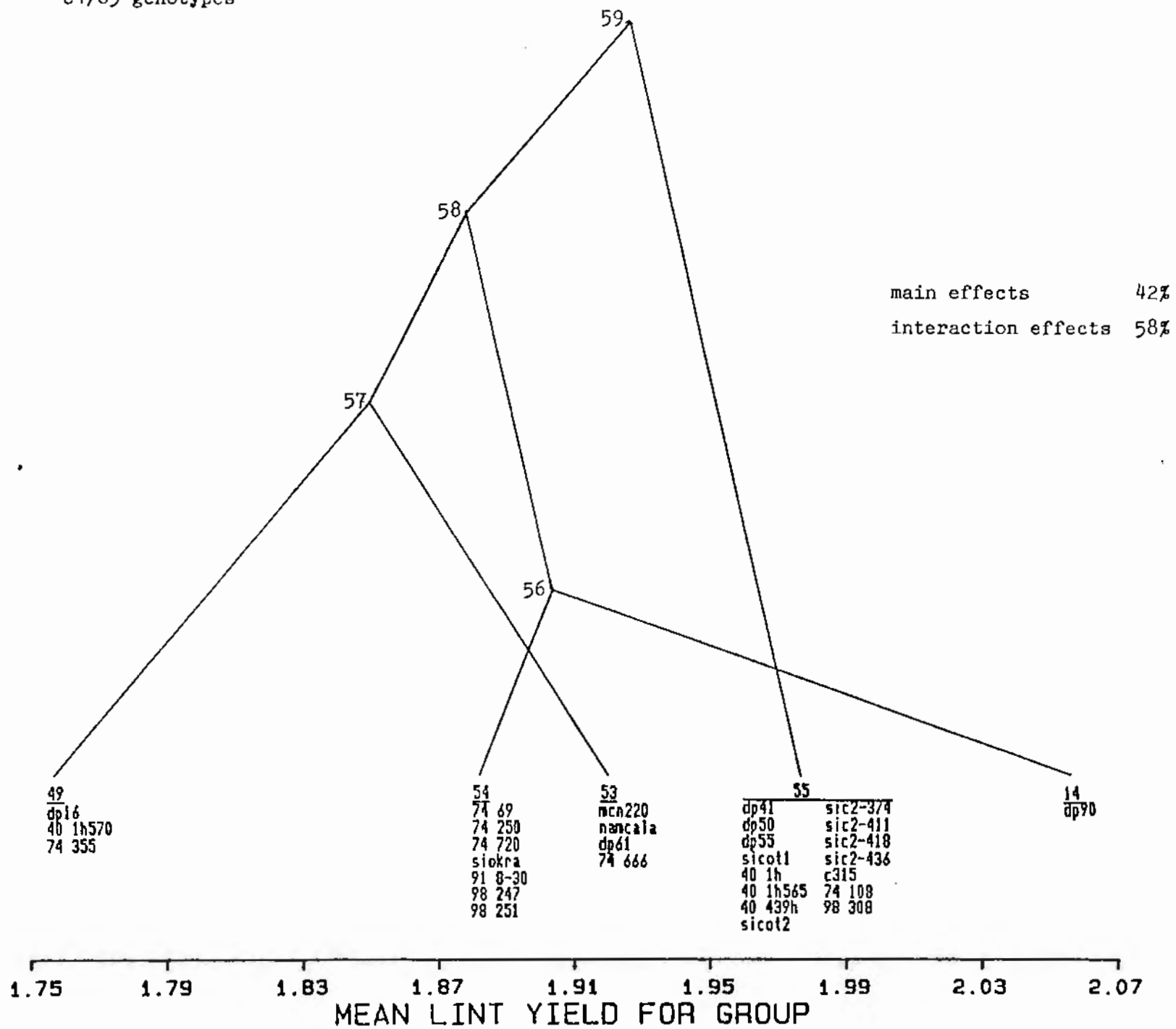
- group 49 - genotypes which are low yielding at all locations.
- group 54 - mostly early maturity genotypes.
 - generally low yielding except darling downs.
- group 53 - mostly USA developed varieties
 - high yielding in central Queensland.
- group 55 - mostly deltapine related genotypes.
- group 14 - "Deltapine 90" is genetically different from the other deltapine genotypes.
 - high yielding at most locations.

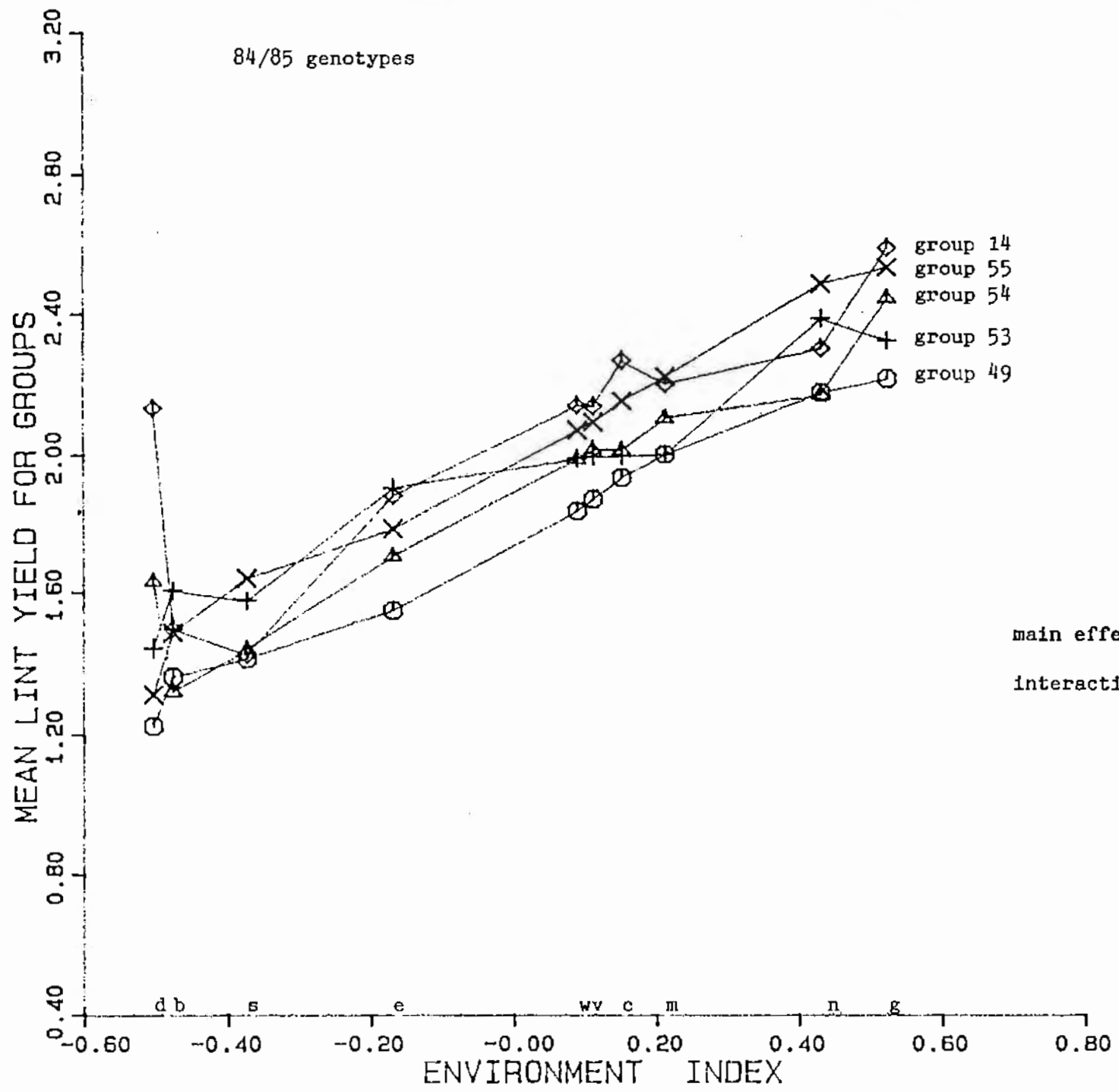
Groups 14 and 54 have similar response patterns over environments, but group 14 is higher yielding than group 54.

Genotype-environment interaction effects (58%) are a major component in separating genotype groups. The main environments contributing to the separation of genotype groups are

- n separates deltapine related genotypes (group 55) from others
- d separates Deltapine 90 from other genotypes
- e separates low yielding genotypes (group 49) from others

84/85 genotypes





main effects 42%

interaction effects 58%

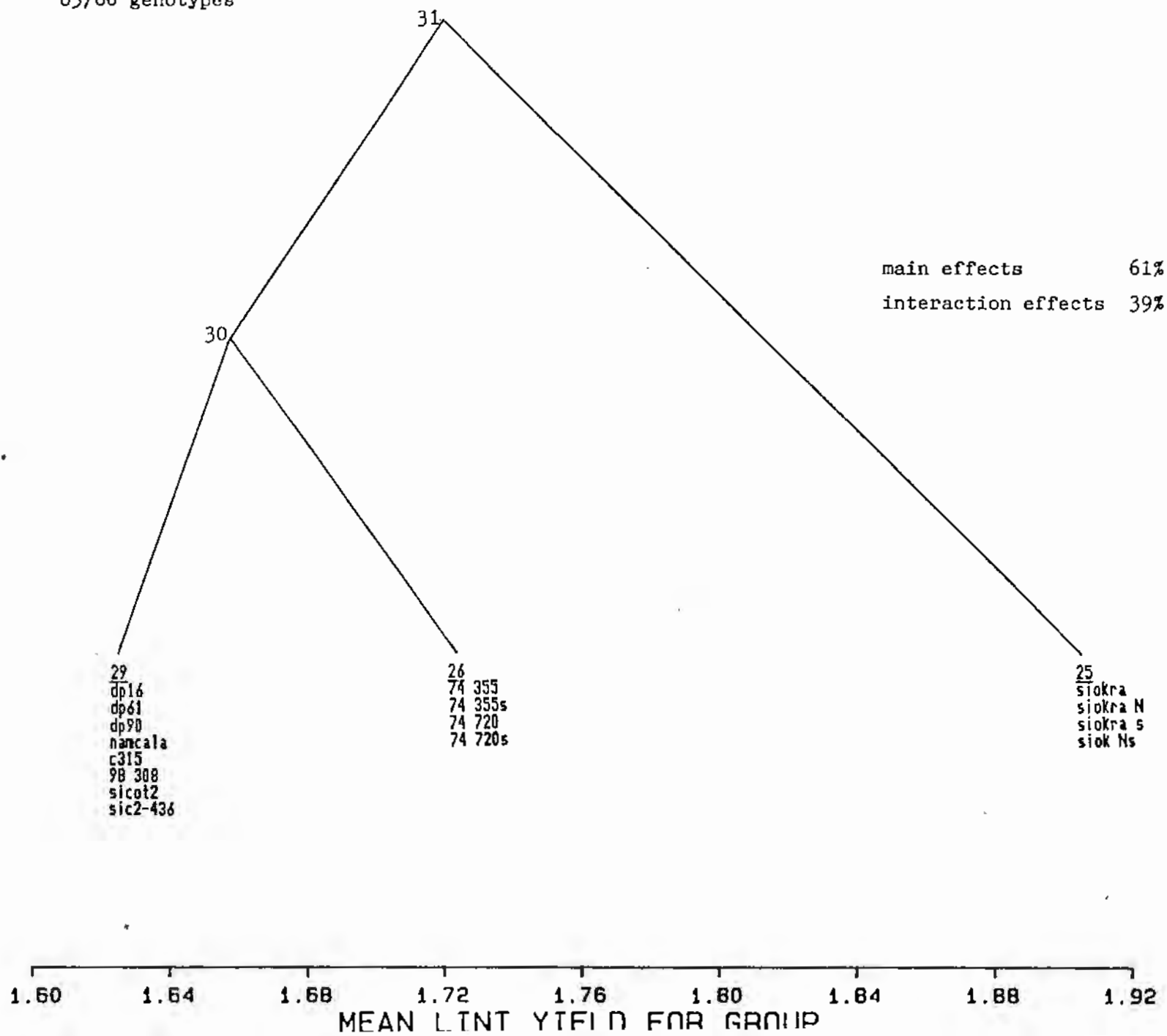
3.12 85/86 genotypes

- group 25 - all recent Siokra selections.
 - highest yielding of the 3 groups, especially at Biloela.
- group 26 - all recent selections from cross n74.
 - intermediate yield of the 3 groups.
- group 29 - remaining genotypes in trial, all older selections.
 - lowest yielding of 3 groups, especially at darling downs.

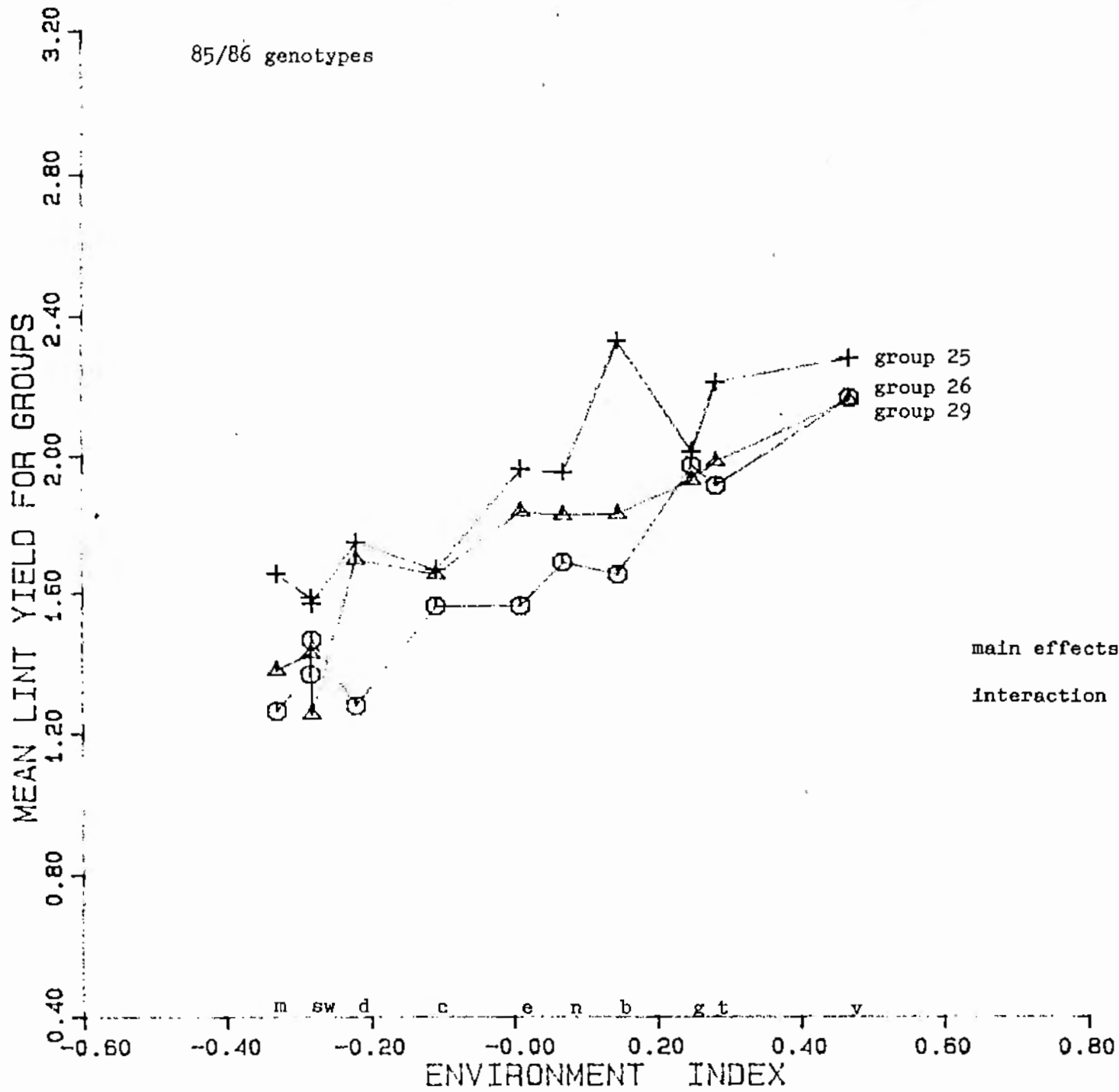
Both genotype main effects (61%) and ge interaction effects (39%) are important in separating genotype groups. The main environments contributing to the separation of genotype groups are

- b separates the Siokra selections (group 25) from the other genotypes.
- d separates the early maturity genotypes (groups 25 and 26) from the other genotypes.

85/86 genotypes



85/86 genotypes



3.13 Summary

On average, ge interaction effects contributed 41% and genotype main effects contributed 59% to the classification of genotypes (Table 3.13, Appendix I). This is clear evidence of the importance of genotype-environment interactions in determining groupings of genotype response patterns over environments.

Table 3.13 Contributions of ge interaction effects (%) to classification of genotypes.

year	ge contribution(%)
74	57
75	26
76	34
77	32
78	45
79	43
80	29
81	51
82	41
83	33
84	58
85	39
average	41

The within group composition of all genotype groups was examined in relation to the genetical origin of the genotypes. In summary, the results indicate that the classification of genotypes into groups is definitely related to the genetical origin of genotypes.

Therefore, the numerical classification analysis is summarizing the genotype-environment interactions in a manner that can easily be interpreted by a cotton breeder and readily used to make decisions about various aspects of a cotton breeding program.

Generalizing over the 12 data sets, the main contribution of locations to the separation of genotype groups is

West namoi	separates high yielding Deltapine related genotypes
Myall vale	similar to west namoi but separation not as significant
Darling downs	separates early maturity genotypes
Biloela	separates some specifically adapted genotypes such as "deltapine gl fr" or namcala related genotypes
Warren	contrasting environment in some years
Emerald	separates generally unadapted genotypes
Theodore	separates Coker related genotypes